

# Package ‘multiWGCNA’

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**Description** An R package for deeping mining gene co-expression networks in multi-trait expression data. Provides functions for analyzing, comparing, and visualizing WGCNA networks across conditions. multiWGCNA was designed to handle the common case where there are multiple biologically meaningful sample traits, such as disease vs wildtype across development or anatomical region.

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multiWGCNA-package     *multiWGCNA: multiWGCNA*

---

### Description

An R package for deeping mining gene co-expression networks in multi-trait expression data. Provides functions for analyzing, comparing, and visualizing WGCNA networks across conditions. multiWGCNA was designed to handle the common case where there are multiple biologically meaningful sample traits, such as disease vs wildtype across development or anatomical region.

### Author(s)

**Maintainer:** Dario Tommasini <dtommasini@gmail.com> ([ORCID](#))

Authors:

- Brent Fogel [contributor]

---

bidirectionalBestMatches  
*Best matching modules*

---

### Description

Find all the modules from dataset1 that have a best match to a module in dataset2 if that module in dataset2 is also a best match to the module in dataset1

### Usage

```
bidirectionalBestMatches(comparisonList, plot = TRUE)
```

### Arguments

`comparisonList` a list with an elemnt "overlap", which is a data.frame resulting from a call to `computeOverlapsFromWGCNA`

`plot` whether to generate a heatmap; default is TRUE

### Value

A ggplot object

### Author(s)

Dario Tommasini

**Examples**

```

library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
comparisonList = list()
comparisonList$overlaps = computeOverlapsFromWGCNA(astrocyte_networks$EAE, astrocyte_networks$WT)
bidirectionalBestMatches(comparisonList)

```

---

BuildTOMFlowDF

*BuildTOMFlowDF*


---

**Description**

Preprocess for plotting a sankey flow diagram showing the movement of genes from one WGCNA to another WGCNA. Uses the flashClust framework.

**Usage**

```
BuildTOMFlowDF(WGCNAlist, networks, toms, genes_to_label, method = "average")
```

**Arguments**

WGCNAlist	list of WGCNA objects
networks	list of network names of length 2
toms	a list of TOM distance objects of length 2
genes_to_label	genes to label across two networks
method	linkage method to pass to flashClust for clustering, default is average linkage

**Value**

a data.frame

**Author(s)**

Dario Tommasini, Xinye Li

---

cleanDatExpr	<i>cleanDatExpr</i>
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---

**Description**

A function that converts a data.frame where row 1 is gene symbols to a numeric matrix where columns are genes and rows are samples for compatibility with most WGCNA functions.

**Usage**

```
cleanDatExpr(datExpr, checkGenesSamples = FALSE)
```

**Arguments**

datExpr            a data.frame where columns are samples and rows are samples and the gene symbols are in the first row  
checkGenesSamples    call the WGCNA function checkGenesSamples?

**Value**

Returns a datExpr with rows as samples and columns as genes

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_se = eh_query[["EH8223"]]
datExpr = data.frame(X = rownames(assays(astrocyte_se)[[1]]), assays(astrocyte_se)[[1]])
cleanDatExpr(datExpr)
```

---

coexpressionLineGraph	<i>Coexpression Line Graph</i>
-----------------------	--------------------------------

---

**Description**

Plots a line graph showing the co-expression of selected genes across samples

**Usage**

```
coexpressionLineGraph(datExpr, splitBy = 1, fontSize = 2.15, colors = NULL)
```

**Arguments**

datExpr	a data.frame with genes as rows and samples as columns
splitBy	how much to split genes by on line graph
fontSize	the font size of the gene labels
colors	a vector of colors; default is random colors generated by colors function

**Value**

a ggplot object

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
datExpr = GetDatExpr(astrocyte_networks[[1]],
  genes = topNGenes(astrocyte_networks$EAE, "EAE_015", 20))
coexpressionLineGraph(datExpr) +
  geom_vline(xintercept = 20.5, linetype='dashed')
```

---

computeOverlapsFromWGCNA

*computeOverlapsFromWGCNA*

---

**Description**

Computes overlap between the modules of two objects of class WGCNA

**Usage**

```
computeOverlapsFromWGCNA(dataset1, dataset2)
```

**Arguments**

dataset1	an object of class WGCNA to compare with dataset2
dataset2	an object of class WGCNA to compare with dataset1

**Value**

Returns a data.frame showing the overlap results for modules from dataset1 with dataset2

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
computeOverlapsFromWGCNA(astrocyte_networks$EAE, astrocyte_networks$WT)
```

---

constructNetworks	<i>constructNetworks: Construct all the weighted gene correlation networks</i>
-------------------	--

---

**Description**

A high level function that returns all networks possible for a given experimental design

**Usage**

```
constructNetworks(
  datExpr,
  sampleTable,
  conditions1,
  conditions2,
  write = FALSE,
  alphaLevel = 0.05,
  plot = FALSE,
  detectNumbers = TRUE,
  ...
)
```

**Arguments**

datExpr	either a SummarizedExperiment object or data.frame with genes are rows and samples as columns
sampleTable	data.frame with sample names in first column and sample traits in the second and third column. First column should be called "Sample"
conditions1	first design conditions, ie healthy/disease
conditions2	second design conditions, ie frontal lobe/temporal lobe
write	write results out to files?
alphaLevel	significance value passed to findBestTrait function, default is 0.05
plot	plot modules? Default is false
detectNumbers	passed to makeTraitTable2; if you have any numeric traits, make sure this is TRUE so that these get detected.
...	Arguments to pass to blockwiseModules function

**Value**

A list of WGCNA objects, ie level one, two, and three networks.

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
autism_se = eh_query[["EH8219"]]
set.seed(1)
autism_se = autism_se[sample(rownames(autism_se), 500),]
sampleTable = colData(autism_se)
conditions1 = unique(sampleTable[,2])
conditions2 = unique(sampleTable[,3])
autism_networks = constructNetworks(autism_se, sampleTable, conditions1[[1]], conditions2[[1]],
  networkType = "signed", TOMType = "unsigned",
  power = 10, minModuleSize = 100, maxBlockSize = 25000,
  reassignThreshold = 0, minKMEtoStay = 0, mergeCutHeight = 0,
  numericLabels = TRUE, pamRespectsDendro = FALSE,
  deepSplit = 4, verbose = 3)
autism_networks[["combined"]]
```

---

diffCoexpression

*Differential co-expression analysis*

---

**Description**

Performs a differential co-expression analysis given an expression data.frame and a conditions vector

**Usage**

```
diffCoexpression(
  datExpr,
  conditions,
  geneList = NULL,
  plot = FALSE,
  method = c("pearson", "spearman"),
  removeFreeNodes = TRUE,
  labelSize = 0.5,
  labelDist = 0,
  shape = "circle",
  degreeForSize = FALSE,
```

```

    label = FALSE,
    onlyPositive = FALSE,
    z.threshold = NULL,
    FDR.threshold = 0.05,
    nodeSize = 3
  )

```

### Arguments

<code>datExpr</code>	a data.frame containing expression values
<code>conditions</code>	a vector containing conditions for the samples
<code>geneList</code>	vector of genes, will use all genes if NULL (default)
<code>plot</code>	plot a network?
<code>method</code>	either "pearson" or "spearman"
<code>removeFreeNodes</code>	remove free nodes from network?
<code>labelSize</code>	label size
<code>labelDist</code>	distance from labels to nodes
<code>shape</code>	shape of nodes
<code>degreeForSize</code>	should node size correspond to degree?
<code>label</code>	label nodes?
<code>onlyPositive</code>	only draw positive correlations?
<code>z.threshold</code>	z-score threshold
<code>FDR.threshold</code>	FDR threshold
<code>nodeSize</code>	size of node

### Value

A list including a matrix of z-scores, a matrix of raw p-values, a matrix of adjusted p-values, and a summary data.frame

### Author(s)

Dario Tommasini

### Examples

```

library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNadata"))
astrocyte_se = eh_query[["EH8223"]]
datExpr = assays(astrocyte_se)[[1]]
diffCoexpression(datExpr, c(rep(1,20), rep(2,16)),
  geneList = c("Gfap", "Vim", "Aspg", "Serpina3n", "Cp", "Osmr", "Cd44",
    "Cxcl10", "Hspb1", "Timp1", "S1pr3", "Steap4", "Lcn2"))

```

---

diffModuleExpression *Differential module expression*

---

### Description

Runs (and plots) the differential module expression analysis

### Usage

```
diffModuleExpression(  
  WGCNAobject,  
  geneList,  
  design,  
  plotTitle = NULL,  
  mode = c("PC1", "Zscore"),  
  testColumn = 2,  
  refColumn = 3,  
  test = c("ANOVA", "PERMANOVA"),  
  plot = TRUE  
)
```

### Arguments

WGCNAobject	WGCNA object
geneList	vector of genes in WGCNAobject
design	the sampleTable
plotTitle	title for the plot
mode	either PC1 or Zscore, default is PC1
testColumn	the column of the sampleTable to be resolved
refColumn	the column of the sampleTable to be used as biological variation
test	statistical test to perform, either "ANOVA" or "PERMANOVA"
plot	generate a plot?

### Value

a data.frame with the resulting p-values

### Examples

```
library(ExperimentHub)  
eh = ExperimentHub()  
eh_query = query(eh, c("multiWGCNAdata"))  
astrocyte_se = eh_query[["EH8223"]]  
sampleTable = colData(astrocyte_se)  
astrocyte_networks = eh_query[["EH8222"]]
```

```
diffModuleExpression(astrocyte_networks[["combined"]],
  topNGenes(astrocyte_networks$combined, "combined_013"),
  sampleTable,
  test = "ANOVA",
  plotTitle = "combined_013",
  plot = TRUE)
```

---

drawMultiWGCNAnetwork *Draw multiWGCNA network*

---

### Description

Draw a network where nodes are modules and edges represent significant gene overlap. Modules are sorted by levels 1, 2, and 3.

### Usage

```
drawMultiWGCNAnetwork(
  WGCNAlist,
  comparisonList,
  moduleOfInterest,
  design,
  overlapCutoff = 0,
  padjCutoff = 1,
  removeOutliers = TRUE,
  alpha = 1e-50,
  layout = NULL,
  hjust = 0.4,
  vjust = 0.3,
  width = 0.5,
  colors = NULL
)
```

### Arguments

WGCNAlist	list of WGCNA objects
comparisonList	the list of overlap comparisons ie from iterate(myNetworks, overlapComparisons, ...)
moduleOfInterest	module of interest, ie "combined_001"
design	the sampleTable design matrix
overlapCutoff	cutoff to remove module correspondences with less than this number of genes
padjCutoff	cutoff to remove module correspondences above this significance value
removeOutliers	remove outlier modules?
alpha	alpha level of significance

layout	layout of network to be passed to plot function of igraph object, defaults to multiWGCNA custom layout
hjust	horizontal justification of labels
vjust	vertical justification of labels
width	width of labels
colors	colors to use for modules, should be the same length as the number of WGCNA objects in the WGCNAlist. Defaults to random colors for each condition.

**Value**

an igraph plot

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_se = eh_query[["EH8223"]]
sampleTable = colData(astrocyte_se)
astrocyte_networks = eh_query[["EH8222"]]
results = list()
results$overlaps = iterate(astrocyte_networks, overlapComparisons, plot=FALSE)
drawMultiWGCNAnetwork(astrocyte_networks,
  results$overlaps,
  "combined_013",
  sampleTable)
```

---

GetDatExpr

*Get expression data*

---

**Description**

Returns the expression data frame a WGCNA object as a data.frame

**Usage**

```
GetDatExpr(object, genes = NULL)
```

**Arguments**

object	An object of class WGCNA
genes	a list of genes to subset to; default is NULL

**Value**

a data.frame

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
datExpr = GetDatExpr(astrocyte_networks[[1]],
  genes = topNGenes(astrocyte_networks$EAE, "EAE_015", 20))
coexpressionLineGraph(datExpr) +
  geom_vline(xintercept = 20.5, linetype='dashed')
```

---

getModule

*name: Name of WGCNAobject*

---

**Description**

Returns the module name for a given gene

**Usage**

```
getModule(WGCNAobject, gene, use = "dynamicLabels")
```

**Arguments**

WGCNAobject	an object of class WGCNA
gene	a gene symbol, should be present in WGCNA object
use	which module label to use e.g. dynamicLabels or dynamicColors

**Value**

A module name

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
getModule(astrocyte_networks[[1]], 'Srgn')
```

---

getPreservation	<i>getPreservation</i>
-----------------	------------------------

---

**Description**

Performs a network preservation analysis

**Usage**

```
getPreservation(reference, test, nPermutations = 100, write = FALSE)
```

**Arguments**

reference	reference network of class WGCNA
test	test network of class WGCNA
nPermutations	number of permutations to perform; at least 50 permutations
write	write to file?

**Value**

a data.frame summarizing results of preservation analysis

**Author(s)**

Dario Tommasini

---

GetSignificantOverlap	<i>Get significant overlap</i>
-----------------------	--------------------------------

---

**Description**

A function that subsets an overlap comparisons data.frame.

**Usage**

```
GetSignificantOverlap(df, p.adj.threshold = 0.05, overlap.threshold = 10)
```

**Arguments**

df	a data.frame resulting from a call to computeOverlapsFromWGCNA
p.adj.threshold	an object of class WGCNA to compare with dataset2
overlap.threshold	an object of class WGCNA to compare with dataset1

**Value**

Returns a data.frame with significant overlaps

**Author(s)**

Dario Tommasini

---

iterate	<i>iterate: Iterate function across networks</i>
---------	--

---

**Description**

A high level function that iterates functions across a list of WGCNA objects

**Usage**

```
iterate(WGCNAlist, FUN, ...)
```

**Arguments**

WGCNAlist	a vector of objects of type WGCNAobject
FUN	function to iterate, either overlapComparisons or preservationComparisons
...	arguments to be passed on to overlapComparisons or preservationComparisons

**Value**

a comparison list from overlapComparisons or preservationComparisons

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
results = list()
iterate(astrocyte_networks, overlapComparisons, plot=FALSE)
```

---

makeTraitTable	<i>Generate a trait table from a sample table</i>
----------------	---

---

**Description**

Generates a WGCNA-compatible trait table from a sampleTable dataframe. This function is deprecated. Use makeTraitTable2 instead.

**Usage**

```
makeTraitTable(inputTable, column, detectNumbers = FALSE)
```

**Arguments**

inputTable	the sampleTable dataframe
column	the column from the sampleTable to use as traits
detectNumbers	whether to consider traits with numbers as numerical rather than categorical variables

**Value**

a dataframe with integer values denoting the categorical sample traits

**Examples**

```
sampleTable = data.frame(Sample = c(paste0("EAE", 1:10), paste0("WT", 1:10)),
                          Disease = c(rep("EAE", 10), rep("WT", 10)),
                          Region = c(rep(c("Cb1", "Sc"), 5)))
makeTraitTable(sampleTable, 2)
```

---

makeTraitTable2	<i>Generate a trait table from a sample table (version 2)</i>
-----------------	---

---

**Description**

Generates a WGCNA-compatible trait table from a sampleTable dataframe.

**Usage**

```
makeTraitTable2(inputTable, column, detectNumbers = TRUE)
```

**Arguments**

inputTable	the sampleTable data.frame
column	the column from the sampleTable to use as traits
detectNumbers	whether to consider traits with numbers as numerical rather than categorical variables

**Value**

a data.frame with integer values denoting the categorical sample traits

**Examples**

```
sampleTable = data.frame(Sample = c(paste0("EAE", 1:10), paste0("WT", 1:10)),
                        Disease = c(rep("EAE", 10), rep("WT", 10)),
                        Region = c(rep(c("Cb1", "Sc"), 5)))
makeTraitTable2(sampleTable, 2)
```

---

moduleComparisonPlot *Module comparison plot*

---

**Description**

A plotting function that returns a heatmap and barplot for a module

**Usage**

```
moduleComparisonPlot(overlapDf, dataset1, dataset2)
```

**Arguments**

overlapDf	a data.frame resulting from a call to computeOverlapsFromWGCNA
dataset1	an object of class WGCNA to compare with dataset2
dataset2	an object of class WGCNA to compare with dataset1

**Value**

Returns a ggplot object (flowplot and heatmap) showing the module correspondence between two objects of class WGCNA

**Author(s)**

Dario Tommasini

## Examples

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
overlapDf = computeOverlapsFromWGCNA(astrocyte_networks$EAE, astrocyte_networks$WT)
moduleComparisonPlot(overlapDf, astrocyte_networks$EAE, astrocyte_networks$WT)
```

---

moduleExpressionPlot *Plots an expression profile for a module*

---

## Description

A plotting function that returns a heatmap and barplot for a module

## Usage

```
moduleExpressionPlot(
  WGCNAobject,
  geneList,
  mode = c("PC1", "averageZscore"),
  legend = FALSE,
  title = NULL,
  clusterGenes = FALSE
)
```

## Arguments

WGCNAobject	an object of class WGCNAobject
geneList	a vector of gene names to be extracted from WGCNAobject
mode	use first principal component or averageZscore?
legend	plot legend?
title	title of the plot
clusterGenes	cluster heatmap genes by hierarchical clustering?

## Value

a patchworked ggplot object

## Author(s)

Dario Tommasini

## Examples

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
moduleExpressionPlot(astrocyte_networks[["combined"]],
  geneList = topNGenes(astrocyte_networks$combined, "combined_013"))
```

---

ModuleFlowPlot	<i>Module sankey diagram</i>
----------------	------------------------------

---

## Description

A plotting function that returns a plot

## Usage

```
ModuleFlowPlot(
  WGCNAlist,
  comparisonList,
  networks,
  labels = NULL,
  alpha = 1,
  x.scale = 2,
  y.scale = 1,
  width = 0.3,
  height = 0.2,
  color.by = c("trait", "network", "none"),
  color.low = "cyan",
  color.high = "magenta",
  col = NULL,
  my_layout = NULL,
  use.padj = FALSE,
  only.contiguous = TRUE,
  only.signif = TRUE,
  show.legend = TRUE,
  scale.by.size = TRUE,
  spacer = 10,
  label.y = 50,
  label.size = 4,
  base_family = "Helvetica",
  p.adj.threshold = 0.05,
  overlap.threshold = 10
)
```

**Arguments**

<code>WGCNAlist</code>	a data.frame resulting from a call to <code>computeOverlapsFromWGCNA</code>
<code>comparisonList</code>	an object of class <code>WGCNA</code> to compare with dataset2
<code>networks</code>	an object of class <code>WGCNA</code> to compare with dataset1
<code>labels</code>	labels to show under each networks
<code>alpha</code>	alpha value for the node tiles, default is 1
<code>x.scale</code>	x spacing parameter
<code>y.scale</code>	y spacing parameter
<code>width</code>	width of the block
<code>height</code>	height of the block
<code>color.by</code>	color fill for nodes, either 'network', 'trait', or NULL
<code>color.low</code>	default is cyan
<code>color.high</code>	default is magenta
<code>col</code>	color palette for node tiles, default is internal colors function
<code>my_layout</code>	pass a custom layout for nodes? Default is null
<code>use.padj</code>	use p.adj for edge width, default is false (use gene overlap value)
<code>only.contiguous</code>	only show continuous overlaps? Default is TRUE
<code>only.signif</code>	plot only significant overlap?
<code>show.legend</code>	show the legend?
<code>scale.by.size</code>	scale the node height by the size of the module? Default is TRUE.
<code>spacer</code>	space between the nodes
<code>label.y</code>	adjust the y coordinate for the network labels, default is 1
<code>label.size</code>	size of the node labels, default is 4
<code>base_family</code>	font family for ggraph
<code>p.adj.threshold</code>	adjusted p-value significance threshold for overlap
<code>overlap.threshold</code>	threshold for number of genes overlapping

**Value**

Returns a ggalluvial diagram comparing two networks

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
results = list()
results$overlaps = iterate(astrocyte_networks, overlapComparisons, plot=FALSE)
ModuleFlowPlot(astrocyte_networks, results$overlaps, c('combined', 'EAE', 'WT'))
```

---

ModuleFlowPlot2Way      *Module sankey diagram*

---

**Description**

A plotting function that returns a sankey plot comparing two networks. This is a deprecated function, please use ModuleFlowPlot instead.

**Usage**

```
ModuleFlowPlot2Way(
  overlapDf,
  dataset1,
  dataset2,
  only.signif = TRUE,
  show.legend = TRUE,
  col = NULL,
  ...
)
```

**Arguments**

overlapDf	a data.frame resulting from a call to computeOverlapsFromWGCNA
dataset1	an object of class WGCNA to compare with dataset2
dataset2	an object of class WGCNA to compare with dataset1
only.signif	plot only significant overlap?
show.legend	show the legend?
col	color palette, default is internal colors function
...	params to GetSignificantOverlap function for thresholds

**Value**

Returns a ggalluvial diagram comparing two networks

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
overlapDf = computeOverlapsFromWGCNA(astrocyte_networks$EAE, astrocyte_networks$WT)
ModuleFlowPlot2Way(overlapDf, astrocyte_networks$EAE, astrocyte_networks$WT)
```

---

moduleToModuleHeatmap *Module to module heatmap*

---

**Description**

Returns a heatmap where color corresponds to FDR-adjusted overlap (hypergeometric test) and the label corresponds to the number of overlapping genes

**Usage**

```
moduleToModuleHeatmap(
  comparisonDf,
  dataset1 = NULL,
  dataset2 = NULL,
  trait1 = NULL,
  trait2 = NULL,
  list1 = NULL,
  list2 = NULL,
  filterByTrait = FALSE,
  alphaLevel = 0.05
)
```

**Arguments**

comparisonDf	the data.frame output of computeOverlapFromWGCNA
dataset1	optional; WGCNA object for dataset 1
dataset2	optional; WGCNA object for dataset 2
trait1	optional; subset to modules correlated to this trait for dataset 1
trait2	optional; subset to modules correlated to this trait for dataset 2
list1	subset to this list of modules for dataset 1
list2	subset to this list of modules for dataset 2
filterByTrait	only plot for modules that correlate with some trait?
alphaLevel	the alpha level of significance for module-trait correlation, defaults to 0.05

**Value**

A ggplot object

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
overlapDf = computeOverlapsFromWGCNA(astrocyte_networks$EAE, astrocyte_networks$WT)
moduleToModuleHeatmap(overlapDf)
```

---

name	<i>name: Name of WGCNAobject</i>
------	----------------------------------

---

**Description**

Returns the name of a WGCNAobject.

**Usage**

```
name(WGCNAobject)
```

**Arguments**

WGCNAobject     an object of class WGCNA

**Value**

Returns the name of the WGCNA object, ie "EAE" for astrocyte\_networks\$EAE.

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
name(astrocyte_networks$EAE)
```

---

overlapComparisons	<i>Overlap comparisons</i>
--------------------	----------------------------

---

**Description**

Compares modules between two objects of type WGCNAobjects within a WGCNAobject list given the indices. Recommended to be used in conjunction with the iterate function.

**Usage**

```
overlapComparisons(  
  comparisonList,  
  WGCNAlist,  
  first,  
  second,  
  element,  
  plot = TRUE,  
  write = FALSE  
)
```

**Arguments**

comparisonList	a list passed by the iterate function
WGCNAlist	list of objects of class WGCNA
first	index of first WGCNA object
second	index of second WGCNA object
element	element position in the comparison list (passed by iterate function)
plot	generate plots?
write	write results to file?

**Value**

A list, in which the first element is a data.frame showing the overlap results and the second element is a data.frame showing the best matching modules between the two WGCNA objects.

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)  
eh = ExperimentHub()  
eh_query = query(eh, c("multiWGCNAdata"))  
astrocyte_networks = eh_query[["EH8222"]]  
results = list()  
results$overlaps = iterate(astrocyte_networks, overlapComparisons, plot=FALSE)
```

---

performANOVA	<i>Perform ANOVA</i>
--------------	----------------------

---

**Description**

Test association between module expression to traits using ANOVA

**Usage**

```
performANOVA(datExpr, design, testCondition, refCondition, alphaLevel = 0.05)
```

**Arguments**

datExpr	expression data.frame
design	the sampleTable
testCondition	test column in sampleTable
refCondition	reference column in sampleTable
alphaLevel	the significance level

**Value**

a data.frame with p-values for each association

---

permutationTestResults	<i>Permutation test results</i>
------------------------	---------------------------------

---

**Description**

The results of running the PreservationPermutationTest in the astrocyte vignette. This is provided since this function is quite slow. Please see the astrocyte vignette for more details.

**Usage**

```
data(permutationTestResults)
```

**Format**

A list of data.frames containing preservation results for each permutation

---

 PlotMultiNodesTOMflow *PlotMultiNodesTOMflow*


---

### Description

Plots a sankey flow diagram showing the movement of genes from one WGCNA to multi-WGCNA networks. Uses the ggalluvial framework.

### Usage

```
PlotMultiNodesTOMflow(
  TOMDF,
  labels = NULL,
  alpha = 0.1,
  width = 0.05,
  color = "black"
)
```

### Arguments

TOMDF	created by BuildTOMFlowDF
labels	labels to use for the networks, default is Network1, Network2, etc.
alpha	alpha of flows
width	width of the strata
color	color of flows

### Value

a ggplot object

### Author(s)

Dario Tommasini, Xinye Li

---

 preservationComparisonPlot  
*Preservation Comparison Scatterplot*


---

### Description

A plotting function that draws a scatterplot of preservation scores between two WGCNA objects

**Usage**

```
preservationComparisonPlot(  
  preservationList,  
  dataset1,  
  dataset2,  
  alphaLevel = 0.05,  
  outliers = FALSE  
)
```

**Arguments**

<code>preservationList</code>	a list resulting from a call to <code>preservationComparisons</code>
<code>dataset1</code>	an object of class <code>WGCNAobject</code> to compare with <code>dataset2</code>
<code>dataset2</code>	an object of class <code>WGCNAobject</code> to compare with <code>dataset1</code>
<code>alphaLevel</code>	alpha level of significance, default is 0.05
<code>outliers</code>	leave outlier modules? By default these are removed

**Value**

a `ggplot` object

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)  
eh = ExperimentHub()  
eh_query = query(eh, c("multiWGCNAdata"))  
astrocyte_networks = eh_query[["EH8222"]]  
results = list()  
results$preservation=iterate(astrocyte_networks[c("EAE", "WT")],  
  preservationComparisons,  
  write=FALSE,  
  plot=FALSE,  
  nPermutations=2)  
preservationComparisonPlot(results$preservation$EAE_vs_WT,  
  astrocyte_networks$EAE,  
  astrocyte_networks$WT)
```

---

preservationComparisons

*Preservation comparisons*

---

### Description

A high level function that performs a perservation comparison between two WGCNAobjects in a WGCNAlist, usually supplied by iterate function

### Usage

```
preservationComparisons(  
  comparisonList,  
  WGCNAlist,  
  first,  
  second,  
  element,  
  plot = FALSE,  
  write = FALSE,  
  alphaLevel = 0.05,  
  nPermutations = 100  
)
```

### Arguments

comparisonList	a list passed by the iterate function
WGCNAlist	list of objects of type WGCNAobject
first	index of first WGCNAobject
second	index of second WGCNAobject
element	element position in the comparison list (passed by iterate function)
plot	generate plots?
write	write results to file?
alphaLevel	alpha level of significance for module-trait correlation
nPermutations	number of permutations, defaults to 100

### Value

a list of preservation comparisons results across levels 1, 2, 3

### Author(s)

Dario Tommasini

**Examples**

```

library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
results = list()
iterate(astrocyte_networks[c("EAE", "WT")],
  preservationComparisons,
  write=FALSE,
  plot=FALSE,
  nPermutations=2)

```

---

 PreservationPermutationTest

*PreservationPermutationTest*


---

**Description**

Performs a permutation test to determine if a null distribution of expected preservation scores for modules in this dataset if the labels were randomly assigned. Please look at the astrocyte vignette for more info.

**Usage**

```

PreservationPermutationTest(
  referenceDatExpr,
  design,
  constructNetworksIn,
  testPreservationIn,
  nPermutations = 100,
  nPresPermutations = 100,
  ...
)

```

**Arguments**

referenceDatExpr	
	the combined datExpr
design	the sampleTable
constructNetworksIn	
	the condition to use for network construction, e.g. for the astrocyte data, this is "EAE"
testPreservationIn	
	the condition to use for testing preservation, e.g. for the astrocyte data, this was "WT"
nPermutations	the number of permutations to perform for permutation test

```
nPresPermutations      the number of permutations to perform in modulePreservation function
...                    arguments to pass to blockwiseModules function for network construction (should
                        be the same as used for constructing the original network)
```

**Value**

A list of data.frames with preservation results for each permutation

**Author(s)**

Dario Tommasini

**Examples**

```
## Not run:
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
astrocyte_se = eh_query[["EH8223"]]
sampleTable = colData(astrocyte_se)
results = list()
results$permutation.test = PreservationPermutationTest(
  astrocyte_networks$combined@datExpr[sample(17000,3000),],
  sampleTable,
  constructNetworksIn = "EAE", # Construct networks using EAE samples
  testPreservationIn = "WT", # Test preservation of disease samples in WT
  nPermutations = 10, # Number of permutations for permutation test
  nPresPermutations = 10, # Number of permutations for modulePreservation
  networkType = "signed", TOMType = "unsigned",
  power = 12, minModuleSize = 100, maxBlockSize = 25000,
  reassignThreshold = 0, minKMEtoStay = 0, mergeCutHeight = 0,
  numericLabels = TRUE, pamRespectsDendro = FALSE,
  deepSplit = 4, verbose = 3
)

## End(Not run)
```

---

PreservationScoreDistribution

*PreservationScoreDistribution*

---

**Description**

Extracts the preservation score distribution from the results of PreservationPermutationTest.

**Usage**

```
PreservationScoreDistribution(preservationData, moduleOfInterestSize)
```

**Arguments**

preservationData  
the results from PreservationPermutationTest  
moduleOfInterestSize  
the number of genes in your module of interest

**Value**

A data.frame with Z-summary preservation scores of the module from each permutation and the corresponding module size

**Author(s)**

Dario Tommasini

**Examples**

```
# Remove outlier modules
permutationTestResultsFiltered = lapply(permutationTestResults, function(x)
x[!x$is.outlier.module,])

# Find preservation score distribution for a given module size
scores.summary = PreservationScoreDistribution(
permutationTestResultsFiltered,
moduleOfInterestSize = 303)
```

---

runDME

*Run differential module expression*

---

**Description**

A wrapper to run diffModuleExpression on all the modules in a network

**Usage**

```
runDME(  
  WGCNAobject,  
  design,  
  alphaLevel = 0.05,  
  testCondition = NULL,  
  refCondition = NULL,  
  p.adjust = "fdr",  
  plot = FALSE,  
  test = c("ANOVA", "PERMANOVA"),  
  write = FALSE,  
  out = NULL  
)
```

**Arguments**

WGCNAobject	object of class WGCNA with the modules to run DME on
design	the sampleTable
alphaLevel	level of significance
testCondition	the column of the sampleTable to be resolved
refCondition	the column of the sampleTable to be used as biological variation
p.adjust	adjust for multiple comparisons, argument to pass to p.adjust function
plot	generate a plot?
test	statistical test to perform, either "ANOVA" or "PERMANOVA"
write	write results to a file?
out	file name for DME plots, only used if write is TRUE

**Value**

a data.frame summarizing the results of the analysis

**Author(s)**

Dario Tommasini

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_se = eh_query[["EH8223"]]
sampleTable = colData(astrocyte_se)
astrocyte_networks = eh_query[["EH8222"]]
runDME(astrocyte_networks[["combined"]],
      design = sampleTable,
      p.adjust = "fdr",
      refCondition = "Region",
      testCondition = "Disease")
```

---

summarizeResults

*summarizeResults: Summarize results from a results list object*

---

**Description**

Prints (or writes) a summary of the results from a results list object

**Usage**

```
summarizeResults(  
  myNetworks,  
  results,  
  alphaLevel = 0.05,  
  write = FALSE,  
  outputFile = "results.txt"  
)
```

**Arguments**

myNetworks	a list of WGCNAobjects
results	results list
alphaLevel	alpha level of significance
write	write to file?
outputFile	name of output file, defaults to results.txt

**Value**

prints a summary of results from the multiWGCNA analysis

---

TOMFlowPlot

*TOMFlowPlot*

---

**Description**

Plots a sankey flow diagram showing the movement of genes from one WGCNA to another WGCNA. Uses the ggalluvial framework.

**Usage**

```
TOMFlowPlot(  
  WGCNAlist,  
  networks,  
  toms,  
  genes_to_label,  
  alpha = 0.1,  
  color = "black",  
  width = 0.05  
)
```

**Arguments**

WGCNAlist	list of WGCNA objects
networks	list of network names of length 2
toms	a list of TOM distance objects of length 2
genes_to_label	genes to label across two networks
alpha	alpha of flows
color	color of flows
width	width of the strata

**Value**

a ggplot object

**Author(s)**

Dario Tommasini

---

topNGenes	<i>topNGenes: Top N genes of a module</i>
-----------	---

---

**Description**

Returns the top N number of genes of a module. All genes returned if no number is specified. Genes are in order of intramodular connectivity.

**Usage**

```
topNGenes(WGCNAobject, module, nGenes = NULL)
```

**Arguments**

WGCNAobject	an object of class WGCNA
module	the name of the module in WGCNAobject
nGenes	an integer from 1 to module size; returns all genes if left NULL

**Value**

a character vector of the genes/probes in the module

**Examples**

```
library(ExperimentHub)
eh = ExperimentHub()
eh_query = query(eh, c("multiWGCNAdata"))
astrocyte_networks = eh_query[["EH8222"]]
topNGenes(astrocyte_networks$EAE, "EAE_015", nGenes = 10)
```

---

`WGCNA-class`*The WGCNA Class*

---

**Description**

The WGCNA class is the main class used in multiWGCNA to store results from a weighted gene co-expression network analysis. These include the original unaltered expression data used as input, connectivity metrics, module assignment, input sample conditions, trait

**Value**

NA

**Slots**

`datExpr` The expression data, connectivity data, and module assignment

`conditions` A data.frame with integer conditions for WGCNA

`trait` A data.frame showing pearson correlation values to traits

`moduleEigengenes` A data.frame of module eigengenes for each module across samples

`outlierModules` A vector of modules classified by our algorithm as being driven by sample outliers

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