

Package ‘MSstatsConvert’

May 26, 2026

Title Import Data from Various Mass Spectrometry Signal Processing
Tools to MSstats Format

Version 1.22.0

Description

MSstatsConvert provides tools for importing reports of Mass Spectrometry data processing tools into R format suitable for statistical analysis using the MSstats and MSstatsTMT packages.

License Artistic-2.0

Encoding UTF-8

LazyData true

Roxygen list(markdown = TRUE)

RoxygenNote 7.3.3

biocViews MassSpectrometry, Proteomics, Software, DataImport,
QualityControl

Depends R (>= 4.0)

Imports data.table, log4r, methods, checkmate, utils, stringi, Rcpp,
parallel

Suggests tinytest, covr, knitr, arrow, rmarkdown

LinkingTo Rcpp

Collate 'clean_ProteinProspector.R' 'clean_Metamorpheus.R'
'clean_DIANN.R' 'clean_Philosopher.R' 'clean_Spectronaut.R'
'clean_SpectroMine.R' 'clean_Skyline.R'
'clean_ProteomeDiscoverer.R' 'clean_Progenesis.R'
'clean_OpenSWATH.R' 'clean_OpenMS.R' 'clean_MaxQuant.R'
'clean_DIAUmpire.R' 'MSstatsConvert_core_functions.R'
'RcppExports.R' 'converters_DIANNtoMSstatsFormat.R'
'converters_DIAUmpiretoMSstatsFormat.R'
'converters_FragPipetoMSstatsFormat.R'
'converters_MaxQtoMSstatsFormat.R'
'converters_MaxQtoMSstatsTMTFormat.R'
'converters_MetamorpheusToMSstatsFormat.R'
'converters_OpenMStoMSstatsFormat.R'
'converters_OpenMStoMSstatsTMTFormat.R'

'converters_OpenSWATHtoMSstatsFormat.R'
 'converters_PDtoMSstatsFormat.R'
 'converters_PDtoMSstatsTMTFormat.R'
 'converters_PhilosophertoMSstatsTMTFormat.R'
 'converters_ProgenisistoMSstatsFormat.R'
 'converters_ProteinProspectortoMSstatsTMTFormat.R'
 'converters_SkylinetoMSstatsFormat.R'
 'converters_SpectroMinetoMSstatsTMTFormat.R'
 'converters_SpectronauttoMSstatsFormat.R'
 'utils_MSstatsConvert.R' 'utils_annotation.R'
 'utils_anomaly_score.R' 'utils_balanced_design.R'
 'utils_checks.R' 'utils_classes.R' 'utils_clean_features.R'
 'utils_data_health.R' 'utils_documentation.R'
 'utils_dt_operations.R' 'utils_filtering.R' 'utils_fractions.R'
 'utils_logging.R' 'utils_shared_peptides.R'

VignetteBuilder knitr

git_url <https://git.bioconductor.org/packages/MSstatsConvert>

git_branch RELEASE_3_23

git_last_commit 074a559

git_last_commit_date 2026-04-28

Repository Bioconductor 3.23

Date/Publication 2026-05-25

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`.addFractions` *Add a Fraction column to the output of MSstatsPreprocess*

Description

Add a Fraction column to the output of MSstatsPreprocess

Usage

```
.addFractions(input)
```

Arguments

input output of MSstatsPreprocess

Value

data.table

`.adjustIntensities` *Fix invalid intensities: infinite to NA, between 0 and 1 to 0*

Description

Fix invalid intensities: infinite to NA, between 0 and 1 to 0

Usage

```
.adjustIntensities(input)
```

Arguments

input data.table

Value

data.table

`.aggregatePSMstoPeptideIons`
Aggregate multiple PSMs to a single peptide ion.

Description

Aggregate multiple PSMs to a single peptide ion.

Usage

```
.aggregatePSMstoPeptideIons(input, feature_columns, summary_function = sum)
```

Arguments

`input` data.table preprocessed by one of the cleanRaw* functions.
`feature_columns` chr, names of columns that define features.
`summary_function` function that will be used to aggregate intensities if needed.

Value

data.table

`.checkAnnotation` *Check if the annotation is valid*

Description

Check if the annotation is valid

Usage

```
.checkAnnotation(input, annotation)
```

Arguments

`input` data processed by the MSstatsClean
`annotation` annotation created by the MSstatsMakeAnnotation function

Value

TRUE invisibly if the annotation is correct, throws an error otherwise

.checkDDA *Check validity of DDA data*

Description

Check validity of DDA data

Usage

.checkDDA(input)

Arguments

input data.table preprocessed by one of the cleanRaw* functions.

Value

logical

logical, TRUE means that the input dataset comes from a DDA experiment

.checkDuplicatedMeasurements
Check if there are duplicated measurements within run

Description

Check if there are duplicated measurements within run

Usage

.checkDuplicatedMeasurements(input)

Arguments

input output of MSstatsPreprocess

Value

character vector of feature labels

.checkMSstatsParams *Check validity of parameters to the MSstatsImport function.*

Description

Check validity of parameters to the MSstatsImport function.

Usage

```
.checkMSstatsParams(  
  input,  
  annotation,  
  feature_columns,  
  remove_shared_peptides,  
  remove_single_feature_proteins,  
  feature_cleaning  
)
```

Value

none, throws an error if any of the assertions fail

.checkMultiRun *Check if fractionation exists*

Description

Check if fractionation exists

Usage

```
.checkMultiRun(input)
```

Arguments

input output of MSstatsPreprocess

Value

list of two elements: `has_fractions` (logical) indicates if fractions was detected in the input dataset, `is_risky` (logical) indicates if there was a problem with detecting fractionation.

`.checkOverlappedFeatures`
Check if any features are measured in multiple fractions

Description

Check if any features are measured in multiple fractions

Usage

```
.checkOverlappedFeatures(input)
```

Arguments

input output of MSstatsPreprocess

Value

data.table

`.cleanByFeature` *Perform by-feature operations.*

Description

Perform by-feature operations.

Usage

```
.cleanByFeature(  
  input,  
  feature_columns,  
  cleaning_control,  
  anomaly_metrics = c()  
)
```

Arguments

input data.table preprocessed by one of the cleanRaw* functions.
feature_columns character vector of names of columns that define features.
cleaning_control named list of two or three elements. See the documentation for MSstatsImport for details.
anomaly_metrics character vector of quality metric column names to be used as features in an anomaly detection model.

Value

data.table

<code>.cleanRawDIANN</code>	<i>Clean raw Diann files</i>
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Description

Clean raw Diann files

Usage

```
.cleanRawDIANN(
  msstats_object,
  MBR = TRUE,
  quantificationColumn = "FragmentQuantCorrected",
  global_qvalue_cutoff = 0.01,
  qvalue_cutoff = 0.01,
  pg_qvalue_cutoff = 0.01,
  calculateAnomalyScores = FALSE,
  anomalyModelFeatures = c(),
  labeledAminoAcids = NULL
)
```

Arguments

`msstats_object` an object of class `MSstatsDIANNFiles`.

`MBR` True if analysis was done with match between runs

`quantificationColumn`

Use 'FragmentQuantCorrected' (default) column for quantified intensities for DIANN 1.8.x. Use 'FragmentQuantRaw' for quantified intensities for DIANN 1.9.x. Use 'auto' for quantified intensities for DIANN 2.x where each fragment intensity is a separate column, e.g. `Fr0Quantity`.

`global_qvalue_cutoff`

The qvalue cutoff for the `Q.Value` column, i.e. the run-specific precursor q-value. Default is 0.01.

`qvalue_cutoff` If `MBR` is false, the qvalue cutoff for the `Global.Q.Value` column, i.e. global precursor q-value. If `MBR` is true, the qvalue cutoff for the `Lib.Q.Value` column, i.e. the q-value for the library created after the first `MBR` pass. Default is 0.01.

`pg_qvalue_cutoff`

If `MBR` is false, the qvalue cutoff for the `Global.PG.Q.Value` column, i.e. the global q-value for the protein group. If `MBR` is true, the qvalue cutoff for the `Lib.PG.Q.Value` column, i.e. the protein group q-value for the library created after the first `MBR` pass. Default is 0.01.

calculateAnomalyScores

Default is FALSE. If TRUE, will run anomaly detection model and calculate anomaly scores for each feature. Used downstream to weigh measurements in differential analysis.

anomalyModelFeatures

character vector of quality metric column names to be used as features in the anomaly detection model. List must not be empty if calculateAnomalyScores=TRUE.

labeledAminoAcids

Character vector of single-letter amino acid codes that carry the SILAC label in protein turnover experiments, e.g. c("K") or c("K", "R"). Supplying this vector opts in to protein-turnover mode; the exact amino acids determine behaviour only in the ModifiedSequence-parsing path described below.

Channel-based path (DIA-NN 2.x exports that include a Channel column): when labeledAminoAcids is non-NULL *and* the input contains a Channel column, Channel values are mapped directly to IsotopeLabelType ("H" → "H", "L" → "L", anything else → NA). The amino acid codes in labeledAminoAcids are **not** used to validate or filter ModifiedSequence in this path.

ModifiedSequence-parsing path (DIA-NN 1.x exports without a Channel column): when labeledAminoAcids is non-NULL and no Channel column is present, each ModifiedSequence is inspected for SILAC suffixes of the form (SILAC-<AA>-H) or (SILAC-<AA>-L), where <AA> is one of the supplied amino acid codes. Matching sequences are classified as "H" or "L"; sequences carrying neither suffix receive IsotopeLabelType = NA. The SILAC suffix is then stripped from PeptideSequence.

When NULL (default), protein-turnover mode is disabled and all peptides receive IsotopeLabelType = "Light".

Value

data.table

.cleanRawDIAUmpire *Clean raw DIAUmpire files*

Description

Clean raw DIAUmpire files

Usage

.cleanRawDIAUmpire(msstats_object, use_frag, use_pept)

Arguments

<code>msstats_object</code>	Object that inherits from <code>MSstatsInputFiles</code> class.
<code>use_frag</code>	TRUE will use the selected fragment for each peptide. 'Selected_fragments' column is required.
<code>use_pept</code>	TRUE will use the selected fragment for each protein 'Selected_peptides' column is required.

Value

data.table

<code>.cleanRawMaxQuant</code>	<i>Clean raw output from MaxQuant</i>
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Description

Clean raw output from MaxQuant

Usage

```
.cleanRawMaxQuant(
  msstats_object,
  protein_id_col,
  remove_by_site = FALSE,
  channel_columns = "Reporterintensitycorrected"
)
```

Arguments

<code>msstats_object</code>	object that inherits from <code>MSstatsInputFiles</code> class.
<code>protein_id_col</code>	character, name of a column with names of proteins.
<code>remove_by_site</code>	logical, if TRUE, proteins only identified by site will be removed.
<code>channel_columns</code>	character, regular expression that identifies channel columns in TMT data.

Value

data.table

.cleanRawMetamorpheus *Clean raw Metamorpheus files*

Description

Clean raw Metamorpheus files

Usage

```
.cleanRawMetamorpheus(msstats_object, MBR = TRUE, qvalue_cutoff = 0.05)
```

Arguments

`msstats_object` an object of class `MSstatsMetamorpheusFiles`.

`MBR` If `TRUE`, the function will include peaks detected by MBR

`qvalue_cutoff` The q-value cutoff for filtering peaks detected by MBR

Value

data.table

.cleanRawOpenMS *Clean raw output from OpenMS*

Description

Clean raw output from OpenMS

Usage

```
.cleanRawOpenMS(msstats_object)
```

Arguments

`msstats_object` an object of class `MSstatsSpectroMineFiles`.

Value

data.table

`.cleanRawOpenSWATH` *Clean raw OpenSWATH files*

Description

Clean raw OpenSWATH files

Usage

```
.cleanRawOpenSWATH(msstats_object)
```

Arguments

`msstats_object` an object of class `MSstatsSpectroMineFiles`.

Value

`data.table`

`.cleanRawPD` *Clean raw Proteome Discoverer data*

Description

Clean raw Proteome Discoverer data

Usage

```
.cleanRawPD(  
  msstats_object,  
  quantification_column,  
  protein_id_column,  
  sequence_column,  
  remove_shared,  
  remove_protein_groups = TRUE,  
  intensity_columns_regexp = "Abundance"  
)
```

Arguments

`msstats_object` an object of class `MSstatsSpectroMineFiles`.

`quantification_column`

chr, name of a column used for quantification.

`protein_id_column`

chr, name of a column with protein IDs.

sequence_column
chr, name of a column with peptide sequences.

remove_shared lgl, if TRUE, shared peptides will be removed.

remove_protein_groups
if TRUE, proteins with numProteins > 1 will be removed.

intensity_columns_regexp
regular expressions that defines intensity columns. Defaults to "Abundance", which means that columns that contain the word "Abundance" will be treated as corresponding to intensities for different channels.

Value

data.table

.cleanRawPDMSstats	<i>Clean raw PD output</i>
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Description

Clean raw PD output

Usage

```
.cleanRawPDMSstats(  
  msstats_object,  
  quantification_column,  
  protein_id_column,  
  sequence_column,  
  remove_shared,  
  run_column = "SpectrumFile"  
)
```

Arguments

msstats_object an object of class MSstatsSpectroMineFiles.

quantification_column
chr, name of a column used for quantification.

protein_id_column
chr, name of a column with protein IDs.

sequence_column
chr, name of a column with peptide sequences.

remove_shared lgl, if TRUE, shared peptides will be removed.

Value

data.table

.cleanRawPDTMT *Clean raw TMT data from Proteome Discoverer*

Description

Clean raw TMT data from Proteome Discoverer

Usage

```
.cleanRawPDTMT(  
  msstats_object,  
  remove_shared = TRUE,  
  remove_protein_groups = TRUE,  
  protein_id_column = "ProteinAccessions",  
  intensity_columns_regexp = "Abundance",  
  run_column = "SpectrumFile"  
)
```

Arguments

msstats_object an object of class *MSstatsSpectroMineFiles*.
remove_shared lgl, if TRUE, shared peptides will be removed.
remove_protein_groups
 if TRUE, proteins with *numProteins* > 1 will be removed.
protein_id_column
 chr, name of a column with protein IDs.
intensity_columns_regexp
 regular expressions that defines intensity columns. Defaults to "Abundance",
 which means that columns that contain the word "Abundance" will be treated as
 corresponding to intensities for different channels.

Value

data.table

.cleanRawPhilosopher *Clean raw Philosopher files*

Description

Clean raw Philosopher files

Usage

```
.cleanRawPhilosopher(  
  msstats_object,  
  protein_id_col,  
  peptide_id_col,  
  channels,  
  remove_shared_peptides  
)
```

Arguments

msstats_object object of class MSstatsPhilosopherFiles
protein_id_col character name of a column that identifies proteins
peptide_id_col character name of a column that identifies peptides
channels character vector of channel labels
remove_shared_peptides logical, if TRUE, shared peptides will be removed based on the IsUnique column from Philosopher output

Value

data.table

.cleanRawProgenesis *Clean raw Progenesis output*

Description

Clean raw Progenesis output

Usage

```
.cleanRawProgenesis(msstats_object, runs, fix_colnames = TRUE)
```

Arguments

msstats_object an object of class MSstatsSpectroMineFiles.
runs chr, vector of Run labels.
fix_colnames lgl, if TRUE, one of the rows will be used as colnames.

Value

data.table

`.cleanRawSkyline` *Clean raw data from Skyline*

Description

Clean raw data from Skyline

Usage

```
.cleanRawSkyline(msstats_object)
```

Arguments

`msstats_object` an object of class `MSstatsSpectroMineFiles`.

Value

`data.table`

`.cleanRawSpectroMineTMT`
Clean raw SpectroMine TMT data

Description

Clean raw SpectroMine TMT data

Usage

```
.cleanRawSpectroMineTMT(msstats_object)
```

Arguments

`msstats_object` an object of class `MSstatsSpectroMineFiles`.

Value

`data.table`

.cleanRawSpectronaut *Clean raw Spectronaut output.*

Description

Clean raw Spectronaut output.

Usage

```
.cleanRawSpectronaut(  
  msstats_object,  
  intensity,  
  calculateAnomalyScores,  
  anomalyModelFeatures,  
  peptideSequenceColumn = "EG.ModifiedSequence",  
  heavyLabels = NULL  
)
```

Arguments

msstats_object an object of class `MSstatsSpectronautFiles`.

intensity Intensity column to use. Accepts legacy enum values 'PeakArea' (default, uses `F.PeakArea`), 'NormalizedPeakArea' (uses `F.NormalizedPeakArea`). Can also be any raw Spectronaut column name passed as a string (e.g. "FG.MS1Quantity"); the column name is standardized internally. For protein turnover workflows the recommended default is "FG.MS1Quantity".

calculateAnomalyScores Default is FALSE. If TRUE, will run anomaly detection model and calculate anomaly scores for each feature. Used downstream to weigh measurements in differential analysis.

anomalyModelFeatures character vector of quality metric column names to be used as features in the anomaly detection model. List must not be empty if `calculateAnomalyScores=TRUE`.

peptideSequenceColumn Name of the Spectronaut column that contains the peptide sequence. Defaults to "EG.ModifiedSequence". The value is standardized internally (dots and spaces removed) before column lookup.

heavyLabels Character list identifying the heavy isotope labels as it appears inside square brackets in the peptide sequence column, e.g. `c("Lys6")` matches peptides containing [Lys6]. `c("Lys6", "Arg10")` matches peptides containing either [Lys6] or [Arg10]. Supports any novel label name reported by Spectronaut (e.g. "Leu6", "Phe10"). When provided, peptides are classified as heavy (`IsotopeLabelType = "H"`), light (`IsotopeLabelType = "L"`), or unlabeled (`IsotopeLabelType = NA`) based on its labeled sequence. When NULL (default) all peptides receive `IsotopeLabelType = "L"`. Useful for protein turnover experiments.

Value

data.table

`.countCommonFeatures` *Get common values from two vectors of features*

Description

Get common values from two vectors of features

Usage

```
.countCommonFeatures(features_1, features_2)
```

Arguments

features_1	vector of feature names
features_2	vector of feature_names

Value

character vector of common values of features_1 and features_2

`.fillValues` *Set column to a single value*

Description

Set column to a single value

Usage

```
.fillValues(input, fill_list)
```

Arguments

input	data.table preprocessed by one of the cleanRaw* functions.
fill_list	named list, names correspond to column names, elements to values that will be used in the columns.

Value

data.table

.filterByPattern *Handle filtering by pattern*

Description

Handle filtering by pattern

Usage

```
.filterByPattern(input, col_name, patterns, filter, drop)
```

Arguments

<code>input</code>	data.table preprocessed by one of the <code>.cleanRaw*</code> functions.
<code>col_name</code>	chr, name of the column with peptide sequences.
<code>patterns</code>	chr, regular expression - matching peptides will be removed from the data.
<code>filter</code>	lgl, if TRUE, peptides will be actually filtered.
<code>drop</code>	lgl, if TRUE, the column will be dropped.

Value

data.table

.filterByScore *Filter PSMs / proteins by a given score column.*

Description

Filter PSMs / proteins by a given score column.

Usage

```
.filterByScore(  
  input,  
  score_column,  
  score_threshold,  
  direction,  
  behavior,  
  handle_na = "keep",  
  fill_value = NA,  
  filter = TRUE,  
  drop = TRUE  
)
```

Arguments

input	data.table preprocessed by one of the <code>.cleanRaw*</code> functions.
score_column	chr, name of the column that contains scores.
score_threshold	num, values below or above this threshold will be removed from the data.
direction	chr, if "greater" only values above the threshold will be retained, if "smaller" - below the threshold.
behavior	chr, if "remove", values below/above the threshold will be removed, if "replace", they will be set to <code>fill_value</code> .
fill_value	if behavior = "replace", values below/above the threshold will be replaced with <code>fill_value</code> . Defaults to NA.
filter	If TRUE, filtering will be performed.
drop	if TRUE, <code>score_column</code> will be removed.

Value

data.table

`.filterExact` *Filter out specified symbols.*

Description

Filter out specified symbols.

Usage

```
.filterExact(
  input,
  col_name,
  filter_symbols,
  behavior,
  fill_value,
  filter,
  drop
)
```

Arguments

input	data.table preprocessed by one of the <code>.cleanRaw*</code> functions.
col_name	chr, name of the column that will be the base for filtering
filter_symbols	character vector of symbols that will be removed
behavior	chr, if "remove", values below/above the threshold will be removed, if "replace", they will be set to <code>fill_value</code> .

fill_value	if behavior = "replace", values below/above the threshold will be replaced with fill_value. Defaults to NA.
filter	lgl, if TRUE, decoy proteins will be removed from the data.
drop	lgl, if TRUE, column that contains decoy proteins will be dropped.

Value

data.table

.filterFewMeasurements
Remove features with a small number of (non-missing) measurements across runs

Description

Remove features with a small number of (non-missing) measurements across runs

Usage

```
.filterFewMeasurements(  
  input,  
  min_intensity,  
  remove_few,  
  feature_columns = NULL  
)
```

Arguments

input	data.table pre-processed by one of the .cleanRaw* functions.
min_intensity	minimum intensity that will be considered non-missing.
remove_few	logical, if TRUE, features that have less than three measurements will be removed. If FALSE, only features with all missing runs will be removed.
feature_columns	chr, vector of names of columns that define features.

Value

data.table

`.filterManyColumns` *Filter rows that contain specified symbols in multiple columns.*

Description

Filter rows that contain specified symbols in multiple columns.

Usage

```
.filterManyColumns(input, filter_columns, filter_symbols)
```

Arguments

`input` `data.table` preprocessed by one of the `cleanRaw*` functions.
`filter_columns` `chr`, names of columns in which elements will be matched and removed.
`filter_symbols` `chr`, vector of strings. Rows with corresponding elements in `filter_columns` will be removed.

Value

`data.table`

`.filterOverlapped` *Remove overlapped features*

Description

Remove overlapped features

Usage

```
.filterOverlapped(input, summary_function, overlapped_features)
```

Arguments

`input` `data.table` preprocessed by one of the `.cleanRaw*` functions and merged with annotation.
`summary_function` summary function (mean, sum, max) that will be used to pick one feature from multiple overlapping features
`overlapped_features` features that overlap.

Value

`data.table`

.findAvailable *Select an available options from a set of possibilities*

Description

Select an available options from a set of possibilities

Usage

```
.findAvailable(possibilities, option_set, fall_back = NULL)
```

Arguments

possibilities possible legal values of a variable
option_set set of values that includes one of the possibilities
fall_back if there is none of the possibilities in option_set, or there are multiple hits,
 default to fall_back

Value

same as option_set, usually character

.fixBasicColumns *Remove underscores from sequences and change intensity type to numeric*

Description

Remove underscores from sequences and change intensity type to numeric

Usage

```
.fixBasicColumns(input)
```

Arguments

input data.table

Value

data.table

`.fixColumnTypes` *Change classes of multiple columns*

Description

Change classes of multiple columns

Usage

```
.fixColumnTypes(
  input,
  numeric_columns = NULL,
  character_columns = NULL,
  factor_columns = NULL
)
```

Arguments

`input` data.table preprocessed by one of the `cleanRaw*` functions.
`numeric_columns` chr, vector of names of columns that will be converted to numeric.
`character_columns` chr, vector of names of columns that will be converted to character.
`factor_columns` chr, vector of names of columns that will be converted to factor.

Value

data.table

`.fixMissingValues` *Change labels for missing values*

Description

Change labels for missing values

Usage

```
.fixMissingValues(input, fix_missing = NULL)
```

Arguments

`input` output of `MSstatsPreprocess`
`fix_missing` missing values can be labeled by `NA`, `0` or both. If `NULL`, data were processed by `Skyline`, so missing values will be denoted by both `NA` and `0`. If `"na_to_zero"`, `NA` values will be replaced by `0`. If `"zero_to_na"`, `0` values will be replaced by `NA`

Value

data.table

.getChannelColumns	<i>Get intensity columns from wide-format data</i>
--------------------	--

Description

Get intensity columns from wide-format data

Usage

```
.getChannelColumns(col_names, ...)
```

Arguments

- col_names names of columns, where some of the columns store intensity value for different channels
- ... varying number of strings that define channel columns.

Value

character vector of column names that correspond to channel intensities

.getDataTable	<i>Read file from a provided path or convert given data.frame to data.table</i>
---------------	---

Description

Read file from a provided path or convert given data.frame to data.table

Usage

```
.getDataTable(input, ...)
```

Arguments

- input report from a signal processing tool or a path to it
- ... additional parameters for data.table::fread

Value

data.table

`.getFullDesign` *Create a data.frame of each combination of values for given variables*

Description

Create a data.frame of each combination of values for given variables

Usage

```
.getFullDesign(input, group_col, feature_col, measurement_col, is_tmt)
```

Arguments

<code>input</code>	output of MSstatsPreprocess
<code>group_col</code>	name of column in input. Combination of values of <code>feature_col</code> and <code>measurement_col</code> will be created within each unique value of this column
<code>is_tmt</code>	if TRUE, data will be treated as coming from TMT experiment.
<code>'feature_col'</code>	name of the column that labels features
<code>'measurement_col'</code>	name of a column with measurement labels - Runs in label-free case, Channels in TMT case.

Value

data.table

`.getMissingRunsPerFeature`
Get names of missing runs

Description

Get names of missing runs

Usage

```
.getMissingRunsPerFeature(input)
```

Arguments

<code>input</code>	output of MSstatsPreprocess
--------------------	-----------------------------

Value

data.table

.getOverlappingFeatures

Get features that are overlapped among multiple runs

Description

Get features that are overlapped among multiple runs

Usage

```
.getOverlappingFeatures(input)
```

Arguments

input	data.table preprocessed by one of the .cleanRaw* functions and merged with annotation.
-------	--

Value

data.table

.getPhilosopherInput *Convert Philosopher parameters to consistent format*

Description

Convert Philosopher parameters to consistent format

Usage

```
.getPhilosopherInput(input, path, folder)
```

Arguments

input	data.frame of msstats.csv file produced by Philosopher
path	character. Path to a file or directory containing msstats.csv output(s) from Philosopher. Used when input is NULL.
folder	logical. If TRUE, path is treated as a directory and all msstats files within it are read. If FALSE, path is treated as a single file path.

`.handleFiltering` *Handle PSM/proteins scores*

Description

Handle PSM/proteins scores

Usage

```
.handleFiltering(input, score_filtering, exact_filtering, pattern_filtering)
```

Arguments

`input` `data.table` preprocessed by one of the `.cleanRaw*` functions.
`score_filtering` list of by-score filtering controls.
`exact_filtering` list of exact filtering controls.
`pattern_filtering` list of by-pattern filtering controls.

Value

`data.table`

`.handleFractions` *Check if there are overlapping features and remove if needed*

Description

Check if there are overlapping features and remove if needed

Usage

```
.handleFractions(input)
```

Arguments

`input` `data.table` preprocessed by one of the `.cleanRaw*` functions and merged with annotation.

Value

`data.table`

.handleFractionsLF *Handle overlapping features*

Description

Handle overlapping features

Usage

`.handleFractionsLF(input)`

Arguments

input output of MSstatsPreprocess

Value

data.table

.handleFractionsTMT *Remove peptide ions overlapped among multiple fractions of the same biological mixture*

Description

Remove peptide ions overlapped among multiple fractions of the same biological mixture

Usage

`.handleFractionsTMT(input)`

Arguments

input data.table preprocessed by one of the `.cleanRaw*` functions and merged with annotation.

Value

data.table

`.handleIsotopicPeaks` *Handle isotopic peaks*

Description

Handle isotopic peaks

Usage

```
.handleIsotopicPeaks(input, aggregate = FALSE)
```

Arguments

`input` data.table preprocessed by one of the `cleanRaw*` functions.
`aggregate` if TRUE, isotopic peaks will be summed.

Value

data.table

`.handleSharedPeptides` *Handle shared peptides.*

Description

Handle shared peptides.

Usage

```
.handleSharedPeptides(  
  input,  
  remove_shared = TRUE,  
  protein_column = "ProteinName",  
  peptide_column = "PeptideSequence"  
)
```

Arguments

`input` data.table pre-processed by one of the `.cleanRaw*` functions.
`remove_shared` lgl, if TRUE, shared peptides will be removed
`protein_column` chr, name of the column with names of proteins.
`peptide_column` chr, name of the column with peptide sequences.

Value

data.table

```
.handleSingleFeaturePerProtein
```

Remove proteins only identified by a single feature

Description

Remove proteins only identified by a single feature

Usage

```
.handleSingleFeaturePerProtein(input, remove_single_feature)
```

Arguments

input data.table pre-processed by one of the *.cleanRaw** functions.
remove_single_feature
 lgl, if TRUE, proteins with a single feature will be removed.

Value

data.table

```
.logConverterOptions
```

Log information about converter options

Description

Log information about converter options

Usage

```
.logConverterOptions(  
  feature_columns,  
  remove_shared_peptides,  
  remove_single_feature_proteins,  
  feature_cleaning,  
  is_tmt = FALSE  
)
```

Arguments

feature_columns	character vector of names of columns that define spectral features.
remove_shared_peptides	logical, if TRUE shared peptides will be removed.
remove_single_feature_proteins	logical, if TRUE, proteins that only have one feature will be removed.
feature_cleaning	named list with maximum two (for MSstats converters) or three (for MSstatsTMT converter) elements. If handle_few_measurements is set to "remove", feature with less than three measurements will be removed (otherwise it should be equal to "keep"). summarize_multiple_psms is a function that will be used to aggregate multiple feature measurements in a run. It should return a scalar and accept an na.rm parameter. For MSstatsTMT converters, setting remove_psms_with_any_missing will remove features which have missing values in a run from that run.
is_tmt	If TRUE, the dataset comes from a TMT experiment

Value

TRUE invisibly if message was logged

.logSuccess	<i>Make a message about successful data cleaning/importing</i>
-------------	--

Description

Make a message about successful data cleaning/importing

Usage

```
.logSuccess(tool, event)
```

Arguments

tool	name of a signal processing tool
------	----------------------------------

Value

TRUE invisibly if logging was successful

.makeBalancedDesign *Fill missing rows to create balanced design*

Description

Fill missing rows to create balanced design

Usage

```
.makeBalancedDesign(input, fill_missing, anomaly_metrics = c())
```

Arguments

input	output of MSstatsPreprocess
fill_missing	if TRUE, missing Intensities values will be added to data
anomaly_metrics	character vector of quality metric column names to be used as features in an anomaly detection model. and marked as NA

Value

data.table

.makeExactFilterMessage
Make a message about filtering based on fixed values

Description

Make a message about filtering based on fixed values

Usage

```
.makeExactFilterMessage(col_name, filter_symbols, behavior, fill_value)
```

Arguments

col_name	chr, name of the column that will be the base for filtering
filter_symbols	character vector of symbols that will be removed
behavior	chr, if "remove", values below/above the threshold will be removed, if "replace", they will be set to fill_value.
fill_value	if behavior = "replace", values below/above the threshold will be replaced with fill_value. Defaults to NA.

Value

character - message

```
.makeScoreFilterMessage
```

Make a message about filtering based on a score

Description

Make a message about filtering based on a score

Usage

```
.makeScoreFilterMessage(
  score_column,
  score_threshold,
  direction,
  behavior,
  fill_value
)
```

Arguments

<code>score_column</code>	chr, name of the column that contains scores.
<code>score_threshold</code>	num, values below or above this threshold will be removed from the data.
<code>direction</code>	chr, if "greater" only values above the threshold will be retained, if "smaller" - below the threshold.
<code>behavior</code>	chr, if "remove", values below/above the threshold will be removed, if "replace", they will be set to <code>fill_value</code> .
<code>fill_value</code>	if <code>behavior = "replace"</code> , values below/above the threshold will be replaced with <code>fill_value</code> . Defaults to NA.

Value

character - message

```
.mergeAnnotation
```

Merge annotation with feature data

Description

Merge annotation with feature data

Usage

```
.mergeAnnotation(input, annotation)
```

Arguments

annotation data.table with annotation
data.table preprocessed by one of the .cleanRaw functions.

Value

data.table

.MSstatsFormat *Output format for further analysis by MSstats*

Description

Output format for further analysis by MSstats

Usage

```
.MSstatsFormat(input, anomaly_metrics = c())
```

Arguments

input data.table
anomaly_metrics character vector of quality metric column names to be used as features in an anomaly detection model

Value

object of class MSstatsValidated that inherits from data.frame

.nullAppender *log4r appender used not to write messages*

Description

A convenience function written to save time on checking if messages should be printed or logs should be written to a file.

Usage

```
.nullAppender(level, ...)
```

Arguments

level log level
... messages - ignored

Value

NULL invisibly

<code>.onLoad</code>	<i>Set default logging object when package is loaded</i>
----------------------	--

Description

Set default logging object when package is loaded

Usage

```
.onLoad(...)
```

Arguments

`...` ignored

Value

none, sets options called MSstatsLog and MSstatsMsg

<code>.removeOverlappingFeatures</code>	<i>Replace intensities of overlapped fractions with NA, keeping only one fraction</i>
---	---

Description

Replace intensities of overlapped fractions with NA, keeping only one fraction

Usage

```
.removeOverlappingFeatures(input)
```

Arguments

`input` output of MSstatsPreprocess

Value

data.table

.removeSharedPeptides *Remove peptides assigned to more than one protein.*

Description

Remove peptides assigned to more than one protein.

Usage

```
.removeSharedPeptides(input, protein_column, peptide_column)
```

Arguments

input data.table pre-processed by one of the .cleanRaw* functions.
protein_column chr, name of the column with names of proteins.
peptide_column chr, name of the column with peptide sequences.

Value

data.table

.resolveFractionTies *Resolve ties when multiple fractions share the maximum number of measurements for a given feature. In the case of a tie, the fraction with the highest mean intensity is selected.*

Description

Resolve ties when multiple fractions share the maximum number of measurements for a given feature. In the case of a tie, the fraction with the highest mean intensity is selected.

Usage

```
.resolveFractionTies(input, max_fractions)
```

Arguments

input output of MSstatsPreprocess
max_fractions data.table of fractions that share the maximum number of unique runs per feature, as produced by .removeOverlappingFeatures

Value

data.table with columns feature and Fraction, containing one selected fraction per feature

```
.selectMSstatsColumns Select columns for MSstats format
```

Description

Select columns for MSstats format

Usage

```
.selectMSstatsColumns(input, anomaly_metrics)
```

Arguments

input data.table

Value

data.table

```
.sharedParametersAmongConverters  
A dummy function to store shared documentation items for converters.
```

Description

A dummy function to store shared documentation items for converters.

Usage

```
.sharedParametersAmongConverters()
```

Arguments

removeFewMeasurements
TRUE (default) will remove the features that have 1 or 2 measurements across runs.

useUniquePeptide
TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.

summaryForMultipleRows
max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.

removeProtein_with1Feature
TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.

removeProtein_with1Peptide
TRUE will remove the proteins which have only 1 peptide and charge. FALSE is default.

removeOxidationMpeptides
TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.

removeMpeptides
TRUE will remove the peptides including 'M' sequence. FALSE is default.

use_log_file logical. If TRUE, information about data processing will be saved to a file.

append logical. If TRUE, information about data processing will be added to an existing log file.

verbose logical. If TRUE, information about data processing will be printed to the console.

log_file_path character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If append = TRUE, has to be a valid path to a file.

... additional parameters to `data.table::fread`.

.standardizeColnames *Change column names to match read.table/read.csv/read.delim conventions*

Description

Change column names to match read.table/read.csv/read.delim conventions

Usage

```
.standardizeColnames(col_names)
```

Arguments

col_names chr, vector of column names

Value

character vector

```
.summarizeMultipleMeasurements
```

Summarize multiple measurements per feature in a single run

Description

Summarize multiple measurements per feature in a single run

Usage

```
.summarizeMultipleMeasurements(  
  input,  
  aggregator,  
  feature_columns,  
  anomaly_metrics = c()  
)
```

Arguments

<code>input</code>	data.table pre-processed by one of the <code>.cleanRaw*</code> functions.
<code>aggregator</code>	function that will be used to aggregate duplicated values.
<code>feature_columns</code>	chr, vector of names of columns that define features.
<code>anomaly_metrics</code>	character vector of quality metric column names to be used as features in an anomaly detection model.

Value

data.table

```
.summarizeMultiplePSMs
```

Pick one PSM from a data.table of several PSMs.

Description

Pick one PSM from a data.table of several PSMs.

Usage

```
.summarizeMultiplePSMs(input, summary_function)
```

Arguments

input data.table preprocessed by one of the .cleanRaw* functions.
summary_function function that will be used to aggregate intensities if needed.

Value

character - label of a chosen PSM

.validateMSstatsConverterParameters

Generic parameter validation for all MSstats converters using configuration object

Description

Generic parameter validation for all MSstats converters using configuration object

Usage

```
.validateMSstatsConverterParameters(config)
```

Arguments

config A list containing all converter parameters. See details for required structure.

Details

The config list should contain the input and optionally other parameters:

- input: input data (required)
- annotation: annotation data (optional)
- intensity: intensity type (optional)
- filter_with_Qvalue: Q-value filter setting (default: FALSE)
- qvalue_cutoff: Q-value cutoff (default: 0.01)
- useUniquePeptide: unique peptide setting (default: TRUE)
- removeFewMeasurements: remove few measurements setting (default: TRUE)
- removeProtein_with1Feature: remove single feature proteins setting (default: FALSE)
- summaryforMultipleRows: aggregation function (default: max)
- calculateAnomalyScores: anomaly detection setting (default: FALSE)
- anomalyModelFeatures: anomaly model features (default: c())
- anomalyModelFeatureTemporal: temporal features (default: c())
- removeMissingFeatures: missing feature threshold (default: 0.5)

- anomalyModelFeatureCount: feature count for anomaly model (default: 100)
- runOrder: run order data (default: NULL)
- n_trees: number of trees (default: 100)
- max_depth: max tree depth (default: "auto")
- numberOfCores: number of cores (default: 1)
- use_log_file: logging setting (default: TRUE)
- append: append setting (default: FALSE)
- verbose: verbose setting (default: TRUE)
- log_file_path: log file path (default: NULL)
- excludedFromQuantificationFilter: filter setting (default: NULL)

Value

NULL (throws error if validation fails)

as.data.frame.MSstatsValidated

Convert output of converters to data.frame

Description

Convert output of converters to data.frame

Usage

```
## S3 method for class 'MSstatsValidated'  
as.data.frame(x, ...)
```

Arguments

x	object of class MSstatsValidated
...	Additional arguments to be passed to or from other methods.

Value

data.frame

```
as.data.table.MSstatsValidated
  Convert output of converters to data.table
```

Description

Convert output of converters to data.table

Usage

```
## S3 method for class 'MSstatsValidated'
as.data.table(x, ...)
```

Arguments

x object of class MSstatsValidated
 ... Additional arguments to be passed to or from other methods.

Value

data.tables

```
CheckDataHealth        Takes as input the output of the SpectronautoMSstatsFormat function
and calculates various quality metrics to assess the health of the data.
Requires Anomaly Detection model to be fit.
```

Description

Takes as input the output of the SpectronautoMSstatsFormat function and calculates various quality metrics to assess the health of the data. Requires Anomaly Detection model to be fit.

Usage

```
CheckDataHealth(input)
```

Arguments

input MSstats input which is the output of Spectronaut converter

Value

list of two data.tables

DIANNtoMSstatsFormat *Import Diann files*

Description

Import Diann files

Usage

```
DIANNtoMSstatsFormat(
  input,
  annotation = NULL,
  global_qvalue_cutoff = 0.01,
  qvalue_cutoff = 0.01,
  pg_qvalue_cutoff = 0.01,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeOxidationMpeptides = TRUE,
  removeProtein_with1Feature = TRUE,
  MBR = TRUE,
  labeledAminoAcids = NULL,
  quantificationColumn = "FragmentQuantCorrected",
  calculateAnomalyScores = FALSE,
  anomalyModelFeatures = c(),
  anomalyModelFeatureTemporal = c(),
  removeMissingFeatures = 0.5,
  anomalyModelFeatureCount = 100,
  runOrder = NULL,
  n_trees = 100,
  max_depth = "auto",
  numberOfCores = 1,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	name of MSstats input report from Diann, which includes fragment-level data. Output fragment data with <code>-export-quant</code> flag in DIA-NN 2.0
annotation	name of 'annotation.txt' data which includes Condition, BioReplicate, Run.
global_qvalue_cutoff	The qvalue cutoff for the Q.Value column, i.e. the run-specific precursor q-value. Default is 0.01.

- `qvalue_cutoff` If MBR is false, the qvalue cutoff for the Global.Q.Value column, i.e. global precursor q-value. If MBR is true, the qvalue cutoff for the Lib.Q.Value column, i.e. the q-value for the library created after the first MBR pass. Default is 0.01.
- `pg_qvalue_cutoff`
If MBR is false, the qvalue cutoff for the Global.PG.Q.Value column, i.e. the global q-value for the protein group. If MBR is true, the qvalue cutoff for the Lib.PG.Q.Value column, i.e. the protein group q-value for the library created after the first MBR pass. Default is 0.01.
- `useUniquePeptide`
should unique peptides be removed
- `removeFewMeasurements`
should proteins with few measurements be removed
- `removeOxidationMpeptides`
should peptides with oxidation be removed
- `removeProtein_with1Feature`
should proteins with a single feature be removed
- `MBR` True if analysis was done with match between runs
- `labeledAminoAcids`
Character vector of single-letter amino acid codes that carry the SILAC label in protein turnover experiments, e.g. `c("K")` or `c("K", "R")`. Supplying this vector opts in to protein-turnover mode; the exact amino acids determine behaviour only in the ModifiedSequence-parsing path described below.
Channel-based path (DIA-NN 2.x exports that include a Channel column): when `labeledAminoAcids` is non-NULL *and* the input contains a Channel column, Channel values are mapped directly to IsotopeLabelType ("H" → "H", "L" → "L", anything else → NA). The amino acid codes in `labeledAminoAcids` are **not** used to validate or filter ModifiedSequence in this path.
ModifiedSequence-parsing path (DIA-NN 1.x exports without a Channel column): when `labeledAminoAcids` is non-NULL and no Channel column is present, each ModifiedSequence is inspected for SILAC suffixes of the form (SILAC-<AA>-H) or (SILAC-<AA>-L), where <AA> is one of the supplied amino acid codes. Matching sequences are classified as "H" or "L"; sequences carrying neither suffix receive IsotopeLabelType = NA. The SILAC suffix is then stripped from PeptideSequence.
When NULL (default), protein-turnover mode is disabled and all peptides receive IsotopeLabelType = "Light".
- `quantificationColumn`
Use 'FragmentQuantCorrected'(default) column for quantified intensities for DIANN 1.8.x. Use 'FragmentQuantRaw' for quantified intensities for DIANN 1.9.x. Use 'auto' for quantified intensities for DIANN 2.x where each fragment intensity is a separate column, e.g. Fr0Quantity.
- `calculateAnomalyScores`
Default is FALSE. If TRUE, will run anomaly detection model and calculate anomaly scores for each feature. Used downstream to weigh measurements in differential analysis.

<code>anomalyModelFeatures</code>	character vector of quality metric column names to be used as features in the anomaly detection model. List must not be empty if <code>calculateAnomalyScores=TRUE</code> .
<code>anomalyModelFeatureTemporal</code>	character vector of temporal direction corresponding to columns passed to <code>anomalyModelFeatures</code> . Values must be one of: <code>mean_decrease</code> , <code>mean_increase</code> , <code>dispersion_increase</code> , or <code>NULL</code> (to perform no temporal feature engineering). Default is empty vector. If <code>calculateAnomalyScores=TRUE</code> , vector must have as many values as <code>anomalyModelFeatures</code> (even if all <code>NULL</code>).
<code>removeMissingFeatures</code>	Remove features with missing values in more than this fraction of runs. Default is 0.5. Only used if <code>calculateAnomalyScores=TRUE</code> .
<code>anomalyModelFeatureCount</code>	Feature selection for anomaly model. Anomaly detection works on the precursor-level and can be much slower if all features used. We will by default filter to the top-100 highest intensity features. This can be adjusted as necessary. To turn feature-selection off, set this value to a high number (e.g. 10000). Only used if <code>calculateAnomalyScores=TRUE</code> .
<code>runOrder</code>	Temporal order of MS runs. Should be a two column <code>data.table</code> with columns <code>Run</code> and <code>Order</code> , where <code>Run</code> matches the run name output by DIA-NN and <code>Order</code> is an integer. Used to engineer the temporal features defined in <code>anomalyModelFeatureTemporal</code> .
<code>n_trees</code>	Number of trees to use in isolation forest when <code>calculateAnomalyScores=TRUE</code> . Default is 100.
<code>max_depth</code>	Max tree depth to use in isolation forest when <code>calculateAnomalyScores=TRUE</code> . Default is "auto" which calculates depth as $\log_2(N)$ where <code>N</code> is the number of runs. Otherwise must be an integer.
<code>numberOfCores</code>	Number of cores for parallel processing anomaly detection model. When <code>> 1</code> , a logfile named <code>'MSstats_anomaly_model_progress.log'</code> is created to track progress. Only works for Linux & Mac OS. Default is 1.
<code>use_log_file</code>	logical. If <code>TRUE</code> , information about data processing will be saved to a file.
<code>append</code>	logical. If <code>TRUE</code> , information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If <code>TRUE</code> , information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

`data.frame` in the MSstats required format.

Author(s)

Elijah Willie

Examples

```

input_file_path = system.file("tinytest/raw_data/DIANN/diann_input.tsv",
                              package="MSstatsConvert")
annotation_file_path = system.file("tinytest/raw_data/DIANN/annotation.csv",
                                   package = "MSstatsConvert")
input = data.table::fread(input_file_path)
annot = data.table::fread(annotation_file_path)
output = DIANNtoMSstatsFormat(input, annotation = annot, MBR = FALSE,
                              use_log_file = FALSE)

head(output)

# For DIANN 2.0, set quantificationColumn = 'auto'
input_file_path_2_0 = system.file("tinytest/raw_data/DIANN/diann_2.0.parquet",
                                   package="MSstatsConvert")
annotation_file_path_2_0 = system.file("tinytest/raw_data/DIANN/annotation_diann_2.0.csv",
                                       package = "MSstatsConvert")
input_2_0 = arrow::read_parquet(input_file_path_2_0)
annot_2_0 = data.table::fread(annotation_file_path_2_0)
output_2_0 = DIANNtoMSstatsFormat(input_2_0, annotation = annot_2_0, MBR = FALSE,
                                  use_log_file = FALSE, quantificationColumn = 'auto')

head(output_2_0)

```

DIAUmpiretoMSstatsFormat

Import DIA-Umpire files

Description

Import DIA-Umpire files

Usage

```

DIAUmpiretoMSstatsFormat(
  raw.frag,
  raw.pep,
  raw.pro,
  annotation,
  useSelectedFrag = TRUE,
  useSelectedPep = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)

```

Arguments

<code>raw.frag</code>	name of FragSummary_date.xls data, which includes feature-level data.
<code>raw.pep</code>	name of PeptideSummary_date.xls data, which includes selected fragments information.
<code>raw.pro</code>	name of ProteinSummary_date.xls data, which includes selected peptides information.
<code>annotation</code>	name of annotation data which includes Condition, BioReplicate, Run information.
<code>useSelectedFrag</code>	TRUE will use the selected fragment for each peptide. 'Selected_fragments' column is required.
<code>useSelectedPep</code>	TRUE will use the selected peptide for each protein. 'Selected_peptides' column is required.
<code>removeFewMeasurements</code>	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
<code>removeProtein_with1Feature</code>	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
<code>summaryforMultipleRows</code>	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek

Examples

```
diau_frag = system.file("tinytest/raw_data/DIAUmpire/dia_frag.csv",
                        package = "MSstatsConvert")
diau_pept = system.file("tinytest/raw_data/DIAUmpire/dia_pept.csv",
```

```

                                package = "MSstatsConvert")
diau_prot = system.file("tinytest/raw_data/DIAUmpire/diau_prot.csv",
                        package = "MSstatsConvert")
annot = system.file("tinytest/raw_data/DIAUmpire/annot_diau.csv",
                   package = "MSstatsConvert")
diau_frag = data.table::fread(diau_frag)
diau_pept = data.table::fread(diau_pept)
diau_prot = data.table::fread(diau_prot)
annot = data.table::fread(annot)
diau_frag = diau_frag[, lapply(.SD, function(x) if (is.integer(x)) as.numeric(x) else x)]
# In case numeric columns are not interpreted correctly

diau_imported = DIAUmpiretoMSstatsFormat(diau_frag, diau_pept, diau_prot,
                                         annot, use_log_file = FALSE)

head(diau_imported)

```

FragPipetoMSstatsFormat

Import FragPipe files

Description

Import FragPipe files

Usage

```

FragPipetoMSstatsFormat(
  input,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)

```

Arguments

input name of FragPipe msstats.csv export. ProteinName, PeptideSequence, PrecursorCharge, FragmentIon, ProductCharge, IsotopeLabelType, Condition, BioReplicate, Run, Intensity are required.

useUniquePeptide TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.

removeFewMeasurements	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeProtein_with1Feature	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing will be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If append = TRUE, has to be a valid path to a file.
...	additional parameters to <code>data.table::fread</code> .

Value

data.frame in the MSstats required format.

Author(s)

Devon Kohler

Examples

```
fragpipe_raw = system.file("tinytest/raw_data/FragPipe/fragpipe_input.csv",
                           package = "MSstatsConvert")
fragpipe_raw = data.table::fread(fragpipe_raw)
fragpipe_imported = FragPipeToMSstatsFormat(fragpipe_raw, use_log_file = FALSE)
head(fragpipe_imported)
```

getDataTyp

Get type of dataset from an MSstatsInputFiles object.

Description

Get type of dataset from an MSstatsInputFiles object.

Usage

```
getDataType(msstats_object)

## S4 method for signature 'MSstatsInputFiles'
getDataType(msstats_object)
```

Arguments

msstats_object object that inherits from MSstatsInputFiles class.

Value

character - label of a data type. Currently, "MSstats" or "MSstatsTMT"
character "MSstats" or "MSstatsTMT".

Examples

```
evidence_path = system.file("tinytest/raw_data/MaxQuant/mq_ev.csv",
                             package = "MSstatsConvert")
pg_path = system.file("tinytest/raw_data/MaxQuant/mq_pg.csv",
                       package = "MSstatsConvert")
evidence = read.csv(evidence_path)
pg = read.csv(pg_path)
imported = MSstatsImport(list(evidence = evidence, protein_groups = pg),
                          "MSstats", "MaxQuant")
class(imported)
getDataType(imported) # "MSstats"
```

getInputFile	<i>Get one of files contained in an instance of MSstatsInputFiles class.</i>
--------------	--

Description

Get one of files contained in an instance of MSstatsInputFiles class.

Usage

```
getInputFile(msstats_object, file_type)

## S4 method for signature 'MSstatsInputFiles'
getInputFile(msstats_object, file_type = "input")

## S4 method for signature 'MSstatsPhilosopherFiles'
getInputFile(msstats_object, file_type = "input")
```

Arguments

`msstats_object` object that inherits from `MSstatsPhilosopherFiles` class.
`file_type` character name of a type file. Usually equal to "input".

Value

data.table
data.table
data.table

Examples

```
evidence_path = system.file("tinytest/raw_data/MaxQuant/mq_ev.csv",
                             package = "MSstatsConvert")
pg_path = system.file("tinytest/raw_data/MaxQuant/mq_pg.csv",
                      package = "MSstatsConvert")
evidence = read.csv(evidence_path)
pg = read.csv(pg_path)
imported = MSstatsImport(list(evidence = evidence, protein_groups = pg),
                          "MSstats", "MaxQuant")

class(imported)
head(getInputFile(imported, "evidence"))
```

MaxQtoMSstatsFormat *Import MaxQuant files*

Description

Import MaxQuant files

Usage

```
MaxQtoMSstatsFormat(
  evidence,
  annotation,
  proteinGroups,
  proteinID = "Proteins",
  useUniquePeptide = TRUE,
  summaryforMultipleRows = max,
  removeFewMeasurements = TRUE,
  removeMpeptides = FALSE,
  removeOxidationMpeptides = FALSE,
  removeProtein_with1Peptide = FALSE,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
```

```
    ...
  )
```

Arguments

evidence	name of 'evidence.txt' data, which includes feature-level data.
annotation	name of 'annotation.txt' data which includes Raw.file, Condition, BioReplicate, Run, IsotopeLabelType information.
proteinGroups	name of 'proteinGroups.txt' data. It needs to matching protein group ID. If proteinGroups=NULL, use 'Proteins' column in 'evidence.txt'.
proteinID	'Proteins'(default) or 'Leading.razor.protein' for Protein ID.
useUniquePeptide	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
removeFewMeasurements	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeMpeptides	TRUE will remove the peptides including 'M' sequence. FALSE is default.
removeOxidationMpeptides	TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.
removeProtein_with1Peptide	TRUE will remove the proteins which have only 1 peptide and charge. FALSE is default.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing will be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If append = TRUE, has to be a valid path to a file.
...	additional parameters to <code>data.table::fread</code> .

Value

data.frame in the MSstats required format.

Note

Warning: MSstats does not support for metabolic labeling or iTRAQ experiments.

Author(s)

Meena Choi, Olga Vitek.

Examples

```
mq_ev = data.table::fread(system.file("tinytest/raw_data/MaxQuant/mq_ev.csv",
                                     package = "MSstatsConvert"))
mq_pg = data.table::fread(system.file("tinytest/raw_data/MaxQuant/mq_pg.csv",
                                     package = "MSstatsConvert"))
annot = data.table::fread(system.file("tinytest/raw_data/MaxQuant/annotation.csv",
                                     package = "MSstatsConvert"))
maxq_imported = MaxQtoMSstatsFormat(mq_ev, annot, mq_pg, use_log_file = FALSE)
head(maxq_imported)
```

MaxQtoMSstatsTMTFormat

Generate MSstatsTMT required input format from MaxQuant output

Description

Generate MSstatsTMT required input format from MaxQuant output

Usage

```
MaxQtoMSstatsTMTFormat(
  evidence,
  proteinGroups,
  annotation,
  which.proteinid = "Proteins",
  rmProt_Only.identified.by.site = FALSE,
  useUniquePeptide = TRUE,
  rmPSM_withfewMea_withinRun = TRUE,
  rmProtein_with1Feature = FALSE,
  summaryforMultipleRows = sum,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

evidence name of 'evidence.txt' data, which includes feature-level data.
proteinGroups name of 'proteinGroups.txt' data.

annotation	data frame which contains column Run, Fraction, TechRepMixture, Mixture, Channel, BioReplicate, Condition. Refer to the example 'annotation.mq' for the meaning of each column.
which.proteinid	Use 'Proteins' (default) column for protein name. 'Leading.proteins' or 'Leading.razor.proteins' or 'Gene.names' can be used instead to get the protein ID with single protein. However, those can potentially have the shared peptides.
rmProt_Only.identified.by.site	TRUE will remove proteins with '+' in 'Only.identified.by.site' column from proteinGroups.txt, which was identified only by a modification site. FALSE is the default.
useUniquePeptide	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
rmPSM_withfewMea_withinRun	TRUE (default) will remove the features that have 1 or 2 measurements within each Run.
rmProtein_with1Feature	TRUE will remove the proteins which have only 1 peptide and charge. Default is FALSE.
summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing will be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If append = TRUE, has to be a valid path to a file.
...	additional parameters to data.table::fread.

Value

data.frame of class "MSstatsTMT"

Examples

```
evidence = data.table::fread(system.file("tinytest/raw_data/MaxQuantTMT/mq_ev.csv",
                                         package = "MSstatsConvert"))
proteinGroups = data.table::fread(system.file("tinytest/raw_data/MaxQuantTMT/mq_pg.csv",
                                              package = "MSstatsConvert"))
annotation.mq = data.table::fread(system.file("tinytest/raw_data/MaxQuantTMT/mq_annotation.csv",
                                             package = "MSstatsConvert"))
input.mq <- MaxQtoMSstatsTMTFormat(evidence, proteinGroups, annotation.mq)
head(input.mq)
```

```
MetamorpheusToMSstatsFormat
    Import Metamorpheus files
```

Description

Import Metamorpheus files

Usage

```
MetamorpheusToMSstatsFormat(
  input,
  annotation = NULL,
  MBR = TRUE,
  qvalue_cutoff = 0.05,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

<code>input</code>	name of Metamorpheus output file, which is tabular format. Use the AllQuantifiedPeaks.tsv file from the Metamorpheus output.
<code>annotation</code>	name of 'annotation.txt' data which includes Condition, BioReplicate.
<code>MBR</code>	If TRUE, the function will include peaks detected by MBR
<code>qvalue_cutoff</code>	The q-value cutoff for filtering peaks detected by MBR
<code>useUniquePeptide</code>	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
<code>removeFewMeasurements</code>	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
<code>removeProtein_with1Feature</code>	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
<code>summaryforMultipleRows</code>	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.

<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

data.frame in the MSstats required format.

Author(s)

Anthony Wu

Examples

```
input = system.file("tinytest/raw_data/Metamorpheus/QuantifiedPeaks.tsv",
                    package = "MSstatsConvert")
input = data.table::fread(input)
annot = system.file("tinytest/raw_data/Metamorpheus/annotation.csv",
                    package = "MSstatsConvert")
annot = data.table::fread(annot)
metamorpheus_imported = MSstatsConvert:::MetamorpheusToMSstatsFormat(input, annotation = annot)
head(metamorpheus_imported)
```

MSstatsAnomalyScores *Run Anomaly Model*

Description

Detects anomalous measurements in mass spectrometry data using an isolation forest algorithm. This function identifies unusual precursor measurements based on quality metrics and their temporal patterns. For features with insufficient quality metric data, it assigns anomaly scores based on the median score of similar features (same peptide and charge combination). The model supports parallel processing for improved performance on large datasets.

Usage

```
MSstatsAnomalyScores(
  input,
  quality_metrics,
  temporal_direction,
  missing_run_count,
  n_feat,
  run_order,
  n_trees,
  max_depth,
  cores
)
```

Arguments

<code>input</code>	data.table preprocessed by the MSstatsBalancedDesign function
<code>quality_metrics</code>	character vector of quality metrics to use in the model
<code>temporal_direction</code>	character vector of same length as <code>quality_metrics</code> indicating temporal feature to create.
<code>missing_run_count</code>	numeric, maximum allowed fraction of missing runs per feature.
<code>n_feat</code>	numeric, maximum number of features per protein to use in the model.
<code>run_order</code>	data.frame with two columns: Run and Order. Order should be numeric and indicate the order of runs.
<code>n_trees</code>	numeric, number of trees to use in the isolation forest model. Default is 100.
<code>max_depth</code>	numeric or "auto", maximum depth of each tree. Default is "auto" which sets depth to $\log_2(N)$ where N is the number of runs.
<code>cores</code>	numeric, number of cores to use for parallel processing. Default is 1.

Value

data.table

`MSstatsBalancedDesign` *Creates balanced design by removing overlapping fractions and filling incomplete rows*

Description

Creates balanced design by removing overlapping fractions and filling incomplete rows

Usage

```
MSstatsBalancedDesign(
  input,
  feature_columns,
  fill_incomplete = TRUE,
  handle_fractions = TRUE,
  fix_missing = NULL,
  remove_few = TRUE,
  anomaly_metrics = c()
)
```

Arguments

<code>input</code>	data.table processed by the MSstatsPreprocess function
<code>feature_columns</code>	str, names of columns that define spectral features
<code>fill_incomplete</code>	if TRUE (default), ensures that rows with missing data for specific features are added as NA. For example, if the y10 ion of peptideA is measured in the "disease" samples but entirely missing for the "healthy" samples, rows with NA values will be created for the y10 ion of peptideA in the "healthy" group. This process increases the number of rows to account for all possible feature-sample combinations.
<code>handle_fractions</code>	if TRUE (default), overlapping fractions will be resolved
<code>fix_missing</code>	str, optional. Defaults to NULL, which means no action. If not NULL, must be one of the options: "zero_to_na" or "na_to_zero". If "zero_to_na", Intensity values equal exactly to 0 will be converted to NA. If "na_to_zero", missing values will be replaced by zeros.
<code>remove_few</code>	lgl, if TRUE, features with one or two measurements across runs will be removed.
<code>anomaly_metrics</code>	character vector of names of columns with quality metrics

Value

data.frame of class MSstatsValidated

Examples

```
unbalanced_data = system.file("tinytest/raw_data/unbalanced_data.csv",
  package = "MSstatsConvert")
unbalanced_data = data.table::as.data.table(read.csv(unbalanced_data))
balanced = MSstatsBalancedDesign(unbalanced_data,
  c("PeptideSequence", "PrecursorCharge",
    "FragmentIon", "ProductCharge"))
dim(balanced) # Now balanced has additional rows (with Intensity = NA)
# for runs that were not included in the unbalanced_data table
```

MSstatsClean	<i>Clean files generated by a signal processing tools.</i>
--------------	--

Description

Clean files generated by a signal processing tools.

Clean DIAUmpire files

Clean MaxQuant files

Clean OpenMS files

Clean OpenSWATH files

Clean Progenesis files

Clean ProteomeDiscoverer files

Clean Skyline files

Clean SpectroMine files

Clean Spectronaut files

Clean Philosopher files

Clean DIA-NN files

Clean Metamorpheus files

Clean Protein Prospector files

Usage

```
MSstatsClean(msstats_object, ...)
```

```
## S4 method for signature 'MSstatsDIAUmpireFiles'
```

```
MSstatsClean(msstats_object, use_frag, use_pept)
```

```
## S4 method for signature 'MSstatsMaxQuantFiles'
```

```
MSstatsClean(  
  msstats_object,  
  protein_id_col,  
  remove_by_site = FALSE,  
  channel_columns = "Reporterintensitycorrected"  
)
```

```
## S4 method for signature 'MSstatsOpenMSFiles'
```

```
MSstatsClean(msstats_object)
```

```
## S4 method for signature 'MSstatsOpenSWATHFiles'
```

```
MSstatsClean(msstats_object)
```

```
## S4 method for signature 'MSstatsProgenesisFiles'
```

```
MSstatsClean(msstats_object, runs, fix_colnames = TRUE)

## S4 method for signature 'MSstatsProteomeDiscovererFiles'
MSstatsClean(
  msstats_object,
  quantification_column,
  protein_id_column,
  sequence_column,
  remove_shared,
  remove_protein_groups = TRUE,
  intensity_columns_regexp = "Abundance"
)

## S4 method for signature 'MSstatsSkylineFiles'
MSstatsClean(msstats_object)

## S4 method for signature 'MSstatsSpectroMineFiles'
MSstatsClean(msstats_object)

## S4 method for signature 'MSstatsSpectronautFiles'
MSstatsClean(
  msstats_object,
  intensity,
  calculateAnomalyScores,
  anomalyModelFeatures,
  peptideSequenceColumn = "EG.ModifiedSequence",
  heavyLabels = NULL
)

## S4 method for signature 'MSstatsPhilosopherFiles'
MSstatsClean(
  msstats_object,
  protein_id_col,
  peptide_id_col,
  channels,
  remove_shared_peptides
)

## S4 method for signature 'MSstatsDIANNFiles'
MSstatsClean(
  msstats_object,
  MBR = TRUE,
  quantificationColumn = "FragmentQuantCorrected",
  global_qvalue_cutoff = 0.01,
  qvalue_cutoff = 0.01,
  pg_qvalue_cutoff = 0.01,
  calculateAnomalyScores = FALSE,
  anomalyModelFeatures = c(),
)
```

```

    labeledAminoAcids = NULL
  )

## S4 method for signature 'MSstatsMetamorpheusFiles'
MSstatsClean(msstats_object, MBR = TRUE, qvalue_cutoff = 0.05)

## S4 method for signature 'MSstatsProteinProspectorFiles'
MSstatsClean(msstats_object)

```

Arguments

msstats_object object that inherits from MSstatsInputFiles class.

... additional parameter to specific cleaning functions.

use_frag TRUE will use the selected fragment for each peptide. 'Selected_fragments' column is required.

use_pept TRUE will use the selected fragment for each protein 'Selected_peptides' column is required.

protein_id_col character, name of a column with names of proteins.

remove_by_site logical, if TRUE, proteins only identified by site will be removed.

channel_columns character, regular expression that identifies channel columns in TMT data.

runs chr, vector of Run labels.

fix_colnames lgl, if TRUE, one of the rows will be used as colnames.

quantification_column chr, name of a column used for quantification.

protein_id_column chr, name of a column with protein IDs.

sequence_column chr, name of a column with peptide sequences.

remove_shared lgl, if TRUE, shared peptides will be removed.

remove_protein_groups if TRUE, proteins with numProteins > 1 will be removed.

intensity_columns_regexp regular expressions that defines intensity columns. Defaults to "Abundance", which means that columns that contain the word "Abundance" will be treated as corresponding to intensities for different channels.

intensity Intensity column to use. Accepts legacy enum values 'PeakArea' (default, uses F.PeakArea), 'NormalizedPeakArea' (uses F.NormalizedPeakArea). Can also be any raw Spectronaut column name passed as a string (e.g. "FG.MS1Quantity"); the column name is standardized internally. For protein turnover workflows the recommended default is "FG.MS1Quantity".

calculateAnomalyScores Default is FALSE. If TRUE, will run anomaly detection model and calculate anomaly scores for each feature. Used downstream to weigh measurements in differential analysis.

anomalyModelFeatures	character vector of quality metric column names to be used as features in the anomaly detection model. List must not be empty if calculateAnomalyScores=TRUE.
peptideSequenceColumn	Name of the Spectronaut column that contains the peptide sequence. Defaults to "EG.ModifiedSequence". The value is standardized internally (dots and spaces removed) before column lookup.
heavyLabels	Character list identifying the heavy isotope labels as it appears inside square brackets in the peptide sequence column, e.g. c("Lys6") matches peptides containing [Lys6]. c("Lys6", "Arg10") matches peptides containing either [Lys6] or [Arg10]. Supports any novel label name reported by Spectronaut (e.g. "Leu6", "Phe10"). When provided, peptides are classified as heavy (IsotopeLabelType = "H"), light (IsotopeLabelType = "L"), or unlabeled (IsotopeLabelType = NA) based on its labeled sequence. When NULL (default) all peptides receive IsotopeLabelType = "L". Useful for protein turnover experiments.
peptide_id_col	character name of a column that identifies peptides
channels	character vector of channel labels
remove_shared_peptides	logical, if TRUE, shared peptides will be removed based on the IsUnique column from Philosopher output
MBR	True if analysis was done with match between runs
quantificationColumn	Use 'FragmentQuantCorrected'(default) column for quantified intensities for DIANN 1.8.x. Use 'FragmentQuantRaw' for quantified intensities for DIANN 1.9.x. Use 'auto' for quantified intensities for DIANN 2.x where each fragment intensity is a separate column, e.g. Fr0Quantity.
global_qvalue_cutoff	The qvalue cutoff for the Q.Value column, i.e. the run-specific precursor q-value. Default is 0.01.
qvalue_cutoff	If MBR is false, the qvalue cutoff for the Global.Q.Value column, i.e. global precursor q-value. If MBR is true, the qvalue cutoff for the Lib.Q.Value column, i.e. the q-value for the library created after the first MBR pass. Default is 0.01.
pg_qvalue_cutoff	If MBR is false, the qvalue cutoff for the Global.PG.Q.Value column, i.e. the global q-value for the protein group. If MBR is true, the qvalue cutoff for the Lib.PG.Q.Value column, i.e. the protein group q-value for the library created after the first MBR pass. Default is 0.01.
labeledAminoAcids	Character vector of single-letter amino acid codes that carry the SILAC label in protein turnover experiments, e.g. c("K") or c("K", "R"). Supplying this vector opts in to protein-turnover mode; the exact amino acids determine behaviour only in the ModifiedSequence-parsing path described below. Channel-based path (DIA-NN 2.x exports that include a Channel column): when labeledAminoAcids is non-NULL <i>and</i> the input contains a Channel column, Channel values are mapped directly to IsotopeLabelType ("H" → "H",

"L" → "L", anything else → NA). The amino acid codes in labeledAminoAcids are **not** used to validate or filter ModifiedSequence in this path.

ModifiedSequence-parsing path (DIA-NN 1.x exports without a Channel column): when labeledAminoAcids is non-NULL and no Channel column is present, each ModifiedSequence is inspected for SILAC suffixes of the form (SILAC-<AA>-H) or (SILAC-<AA>-L), where <AA> is one of the supplied amino acid codes. Matching sequences are classified as "H" or "L"; sequences carrying neither suffix receive IsotopeLabelType = NA. The SILAC suffix is then stripped from PeptideSequence.

When NULL (default), protein-turnover mode is disabled and all peptides receive IsotopeLabelType = "Light".

Value

data.table
 data.table
 data.table
 data.table
 data.table
 data.table
 data.table
 data.table
 data.table
 data.table
 data.table
 data.table
 data.table
 data.table

Examples

```
evidence_path = system.file("tinytest/raw_data/MaxQuant/mq_ev.csv",
                             package = "MSstatsConvert")
pg_path = system.file("tinytest/raw_data/MaxQuant/mq_pg.csv",
                      package = "MSstatsConvert")
evidence = read.csv(evidence_path)
pg = read.csv(pg_path)
imported = MSstatsImport(list(evidence = evidence, protein_groups = pg),
                          "MSstats", "MaxQuant")
cleaned_data = MSstatsClean(imported, protein_id_col = "Proteins")
head(cleaned_data)
```

MSstatsConvert	<i>MSstatsConvert: An R Package to Convert Data from Mass Spectrometry Signal Processing Tools to MSstats Format</i>
----------------	--

Description

MSstatsConvert helps convert data from different types of mass spectrometry experiments and signal processing tools to a format suitable for statistical analysis with the MSstats and MSstatsTMT packages.

Main functions

[MSstatsLogsSettings](#) for logs management, [MSstatsImport](#) for importing files created by signal processing tools, [MSstatsClean](#) for re-formatting imported files into a consistent format, [MSstatsPreprocess](#) for preprocessing cleaned files, [MSstatsBalancedDesign](#) for handling fractions and creating balanced data.

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MSstatsImport	<i>Import files from signal processing tools.</i>
---------------	---

Description

Import files from signal processing tools.

Usage

```
MSstatsImport(input_files, type, tool, tool_version = NULL, ...)
```

Arguments

<code>input_files</code>	list of paths to input files or data.frame objects. Interpretation of this parameter depends on values of parameters <code>type</code> and <code>tool</code> .
<code>type</code>	chr, "MSstats" or "MSstatsTMT".
<code>tool</code>	chr, name of a signal processing tool that generated input files.
<code>tool_version</code>	not implemented yet. In the future, this parameter will allow handling different versions of each signal processing tools.
<code>...</code>	optional additional parameters to <code>data.table::fread</code> .

Value

an object of class `MSstatsInputFiles`.

Examples

```
evidence_path = system.file("tinytest/raw_data/MaxQuant/mq_ev.csv",
                             package = "MSstatsConvert")
pg_path = system.file("tinytest/raw_data/MaxQuant/mq_pg.csv",
                      package = "MSstatsConvert")
evidence = read.csv(evidence_path)
pg = read.csv(pg_path)
imported = MSstatsImport(list(evidence = evidence, protein_groups = pg),
                          "MSstats", "MaxQuant")
class(imported)
head(getInputFile(imported, "evidence"))
```

MSstatsInputFiles-class

Class to model files that describe a single MS dataset.

Description

Class to model files that describe a single MS dataset.

`MSstatsDIAUmpireFiles`: class for DIAUmpire files.

`MSstatsMaxQuantFiles`: class for MaxQuant files.

`MSstatsOpenMSFiles`: class for OpenMS files.

`MSstatsOpenSWATHFiles`: class for OpenSWATH files.

`MSstatsProgenesisFiles`: class for Progenesis files.

`MSstatsProteomeDiscovererFiles`: class for ProteomeDiscoverer files.

`MSstatsSkylineFiles`: class for Skyline files.

`MSstatsSkylineFiles`: class for SpectroMine files.

`MSstatsSpectronautFiles`: class for Spectronaut files.

MSstatsPhilosopherFiles: class for Philosopher files.
 MSstatsDIANNFiles: class for DIA-NN files.
 MSstatsFragPipeFiles: class for FragPipe files.
 MSstatsMetamorpheusFiles: class for Metamorpheus files.
 MSstatsProteinProspectorFiles: class for ProteinProspector files.

Slots

files named list of files generated by a signal processing tools. In most cases, this will be a single file named input. In some cases, multiple files are used, for example MaxQuant outputs evidence and proteinGroups files.
 type character: "MSstats" or "MSstatsTMT".
 tool character: name of a signal processing tools that generated the output. Possible values are: DIAUmpire, MaxQuant, OpenMS, OpenSWATH, Progenesis, ProteomeDiscoverer, Skyline, SpectroMine, Spectronaut.
 version description of a software version of the signal processing tool. Not implemented yet.

MSstatsLogsSettings *Set how MSstats will log information from data processing*

Description

Set how MSstats will log information from data processing

Usage

```
MSstatsLogsSettings(  
  use_log_file = TRUE,  
  append = FALSE,  
  verbose = TRUE,  
  log_file_path = NULL,  
  base = "MSstats_log_",  
  pkg_name = "MSstats"  
)
```

Arguments

use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing will be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If append = TRUE, has to be a valid path to a file.

base	start of the file name.
pkg_name	currently "MSstats", "MSstatsPTM" or "MSstatsTMT". Each package can use its own separate log settings.

Value

TRUE invisibly in case of successful logging setup.

Examples

```
# No logging and no messages
MSstatsLogsSettings(FALSE, FALSE, FALSE)
# Log, but do not display messages
MSstatsLogsSettings(TRUE, FALSE, FALSE)
# Log to an existing file
file.create("new_log.log")
MSstatsLogsSettings(TRUE, TRUE, log_file_path = "new_log.log")
# Do not log, but display messages
MSstatsLogsSettings(FALSE)
```

MSstatsMakeAnnotation *Create annotation*

Description

Create annotation

Usage

```
MSstatsMakeAnnotation(input, annotation, ...)
```

Arguments

input	data.table preprocessed by the MSstatsClean function
annotation	data.table
...	key-value pairs, where keys are names of columns of annotation

Value

data.table

Examples

```
evidence_path = system.file("tinytest/raw_data/MaxQuant/mq_ev.csv",
                             package = "MSstatsConvert")
pg_path = system.file("tinytest/raw_data/MaxQuant/mq_pg.csv",
                       package = "MSstatsConvert")
evidence = read.csv(evidence_path)
pg = read.csv(pg_path)
imported = MSstatsImport(list(evidence = evidence, protein_groups = pg),
                          "MSstats", "MaxQuant")
cleaned_data = MSstatsClean(imported, protein_id_col = "Proteins")
annot_path = system.file("tinytest/raw_data/MaxQuant/annotation.csv",
                          package = "MSstatsConvert")
mq_annot = MSstatsMakeAnnotation(cleaned_data, read.csv(annot_path),
                                 Run = "Rawfile")
head(mq_annot)
```

MSstatsPreprocess	<i>Preprocess outputs from MS signal processing tools for analysis with MSstats</i>
-------------------	---

Description

Preprocess outputs from MS signal processing tools for analysis with MSstats

Usage

```
MSstatsPreprocess(
  input,
  annotation,
  feature_columns,
  remove_shared_peptides = TRUE,
  remove_single_feature_proteins = TRUE,
  feature_cleaning = list(remove_features_with_few_measurements = TRUE,
                          summarize_multiple_psms = max),
  score_filtering = list(),
  exact_filtering = list(),
  pattern_filtering = list(),
  columns_to_fill = list(),
  aggregate_isotopic = FALSE,
  anomaly_metrics = c(),
  ...
)
```

Arguments

input	data.table processed by the MSstatsClean function.
annotation	annotation file generated by a signal processing tool.

<code>feature_columns</code>	character vector of names of columns that define spectral features.
<code>remove_shared_peptides</code>	logical, if TRUE shared peptides will be removed.
<code>remove_single_feature_proteins</code>	logical, if TRUE, proteins that only have one feature will be removed.
<code>feature_cleaning</code>	named list with maximum two (for MSstats converters) or three (for MSstatsTMT converter) elements. If <code>handle_few_measurements</code> is set to "remove", feature with less than three measurements will be removed (otherwise it should be equal to "keep"). <code>summarize_multiple_psms</code> is a function that will be used to aggregate multiple feature measurements in a run. It should return a scalar and accept an <code>na.rm</code> parameter. For MSstatsTMT converters, setting <code>remove_psms_with_any_missing</code> will remove features which have missing values in a run from that run.
<code>score_filtering</code>	a list of named lists that specify filtering options. Details are provided in the vignette.
<code>exact_filtering</code>	a list of named lists that specify filtering options. Details are provided in the vignette.
<code>pattern_filtering</code>	a list of named lists that specify filtering options. Details are provided in the vignette.
<code>columns_to_fill</code>	a named list of scalars. If provided, columns with names defined by the names of this list and values corresponding to its elements will be added to the output <code>data.frame</code> .
<code>aggregate_isotopic</code>	logical. If TRUE, isotopic peaks will be summed.
<code>anomaly_metrics</code>	character vector of names of columns with quality metrics. Default is missing and is not required if anomaly model not run.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

`data.table`

Examples

```
evidence_path = system.file("tinytest/raw_data/MaxQuant/mq_ev.csv",
                             package = "MSstatsConvert")
pg_path = system.file("tinytest/raw_data/MaxQuant/mq_pg.csv",
                      package = "MSstatsConvert")
evidence = read.csv(evidence_path)
pg = read.csv(pg_path)
imported = MSstatsImport(list(evidence = evidence, protein_groups = pg),
```

```

        "MSstats", "MaxQuant")
cleaned_data = MSstatsClean(imported, protein_id_col = "Proteins")
annot_path = system.file("tinytest/raw_data/MaxQuant/annotation.csv",
                          package = "MSstatsConvert")
mq_annot = MSstatsMakeAnnotation(cleaned_data, read.csv(annot_path),
                                 Run = "Rawfile")

# To filter M-peptides and oxidatin peptides
m_filter = list(col_name = "PeptideSequence", pattern = "M",
                filter = TRUE, drop_column = FALSE)
oxidation_filter = list(col_name = "Modifications", pattern = "Oxidation",
                       filter = TRUE, drop_column = TRUE)
msstats_format = MSstatsPreprocess(
  cleaned_data, mq_annot,
  feature_columns = c("PeptideSequence", "PrecursorCharge"),
  columns_to_fill = list(FragmentIon = NA, ProductCharge = NA),
  pattern_filtering = list(oxidation = oxidation_filter, m = m_filter)
)
# Output in the standard MSstats format
head(msstats_format)

```

MSstatsSaveSessionInfo

Save session information

Description

Save session information

Usage

```

MSstatsSaveSessionInfo(
  path = NULL,
  append = TRUE,
  base = "MSstats_session_info_"
)

```

Arguments

path	optional path to output file. If not provided, "MSstats_session_info" and current timestamp will be used as a file name
append	if TRUE and file given by the path parameter already exists, session info will be appended to the file
base	beginning of a file name

Value

TRUE invisibly after session info was saved

Examples

```
MSstatsSaveSessionInfo("session_info.txt")
MSstatsSaveSessionInfo("session_info.txt", base = "MSstatsTMT_session_info_")
```

OpenMStoMSstatsFormat *Import OpenMS files*

Description

Import OpenMS files

Usage

```
OpenMStoMSstatsFormat(
  input,
  annotation = NULL,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	name of MSstats input report from OpenMS, which includes feature(peptide ion)-level data.
annotation	name of 'annotation.txt' data which includes Condition, BioReplicate, Run. Run should be the same as filename.
useUniquePeptide	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
removeFewMeasurements	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeProtein_with1Feature	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.

<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek.

Examples

```
openms_raw = data.table::fread(system.file("tinytest/raw_data/OpenMS/openms_input.csv",
                                           package = "MSstatsConvert"))
openms_imported = OpenMStoMSstatsFormat(openms_raw, use_log_file = FALSE)
head(openms_imported)
```

OpenMStoMSstatsTMTFormat

Generate MSstatsTMT required input format for OpenMS output

Description

Generate MSstatsTMT required input format for OpenMS output

Usage

```
OpenMStoMSstatsTMTFormat(
  input,
  useUniquePeptide = TRUE,
  rmPSM_withfewMea_withinRun = TRUE,
  rmProtein_with1Feature = FALSE,
  summaryforMultiplePSMs = sum,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

<code>input</code>	MSstatsTMT report from OpenMS
<code>useUniquePeptide</code>	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
<code>rmPSM_withfewMea_withinRun</code>	TRUE (default) will remove the features that have 1 or 2 measurements within each Run.
<code>rmProtein_with1Feature</code>	TRUE will remove the proteins which have only 1 peptide and charge. Default is FALSE.
<code>summaryforMultiplePSMs</code>	sum(default) or max - when there are multiple measurements for certain feature in certain run, select the feature with the largest summation or maximal value.
<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

data.frame of class MSstatsTMT.

Examples

```
raw.om = data.table::fread(system.file("tinytest/raw_data/OpenMSTMT/openmstmt_input.csv",
                                     package = "MSstatsConvert"))
input.om <- OpenMStoMSstatsTMTFormat(raw.om)
head(input.om)
```

OpenSWATHtoMSstatsFormat

Import OpenSWATH files

Description

Import OpenSWATH files

Usage

```

OpenSWATHtoMSstatsFormat(
  input,
  annotation,
  filter_with_mscore = TRUE,
  mscore_cutoff = 0.01,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = max,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)

```

Arguments

<code>input</code>	name of MSstats input report from OpenSWATH, which includes feature-level data.
<code>annotation</code>	name of 'annotation.txt' data which includes Condition, BioReplicate, Run. Run should be the same as filename.
<code>filter_with_mscore</code>	TRUE(default) will filter out the features that have greater than <code>mscore_cutoff</code> in <code>m_score</code> column. Those features will be removed.
<code>mscore_cutoff</code>	Cutoff for <code>m_score</code> . Default is 0.01.
<code>useUniquePeptide</code>	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
<code>removeFewMeasurements</code>	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
<code>removeProtein_with1Feature</code>	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
<code>summaryforMultipleRows</code>	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.

`log_file_path` character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If `append = TRUE`, has to be a valid path to a file.

... additional parameters to `data.table::fread`.

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek.

Examples

```
os_raw = system.file("tinytest/raw_data/OpenSWATH/openswath_input.csv",
                     package = "MSstatsConvert")
annot = system.file("tinytest/raw_data/OpenSWATH/annot_os.csv",
                   package = "MSstatsConvert")
os_raw = data.table::fread(os_raw)
annot = data.table::fread(annot)

os_imported = OpenSWATHtoMSstatsFormat(os_raw, annot, use_log_file = FALSE)
head(os_imported)
```

PDtoMSstatsFormat *Import Proteome Discoverer files*

Description

Import Proteome Discoverer files

Usage

```
PDtoMSstatsFormat(
  input,
  annotation,
  useNumProteinsColumn = FALSE,
  useUniquePeptide = TRUE,
  summaryforMultipleRows = max,
  removeFewMeasurements = TRUE,
  removeOxidationMpeptides = FALSE,
  removeProtein_with1Peptide = FALSE,
  which.quantification = "Precursor.Area",
  which.proteinid = "Protein.Group.Accessions",
  which.sequence = "Sequence",
  use_log_file = TRUE,
```

```

    append = FALSE,
    verbose = TRUE,
    log_file_path = NULL,
    ...
)

```

Arguments

input	PD report or a path to it.
annotation	name of 'annotation.txt' or 'annotation.csv' data which includes Condition, BioReplicate, Run information. 'Run' will be matched with 'Spectrum.File'.
useNumProteinsColumn	TRUE removes peptides which have more than 1 in # Proteins column of PD output.
useUniquePeptide	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
removeFewMeasurements	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeOxidationMpeptides	TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.
removeProtein_with1Peptide	TRUE will remove the proteins which have only 1 peptide and charge. FALSE is default.
which.quantification	Use 'Precursor.Area'(default) column for quantified intensities. 'Intensity' or 'Area' can be used instead.
which.proteinid	Use 'Protein.Accessions'(default) column for protein name. 'Master.Protein.Accessions' can be used instead.
which.sequence	Use 'Sequence'(default) column for peptide sequence. 'Annotated.Sequence' can be used instead.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing will be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If append = TRUE, has to be a valid path to a file.
...	additional parameters to <code>data.table::fread</code> .

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek

Examples

```
pd_raw = system.file("tinytest/raw_data/PD/pd_input.csv",
                     package = "MSstatsConvert")
annot = system.file("tinytest/raw_data/PD/annot_pd.csv",
                   package = "MSstatsConvert")
pd_raw = data.table::fread(pd_raw)
annot = data.table::fread(annot)

pd_imported = PDtoMSstatsFormat(pd_raw, annot, use_log_file = FALSE)
head(pd_imported)
```

PDtoMSstatsTMTFormat *Convert Proteome Discoverer output to MSstatsTMT format.*

Description

Convert Proteome Discoverer output to MSstatsTMT format.

Usage

```
PDtoMSstatsTMTFormat(
  input,
  annotation,
  which.proteinid = "Protein.Accessions",
  useNumProteinsColumn = TRUE,
  useUniquePeptide = TRUE,
  rmPSM_withfewMea_withinRun = TRUE,
  rmProtein_with1Feature = FALSE,
  summaryforMultipleRows = sum,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

<code>input</code>	PD report or a path to it.
<code>annotation</code>	annotation with Run, Fraction, TechRepMixture, Mixture, Channel, BioReplicate, Condition columns or a path to file. Refer to the example 'annotation' for the meaning of each column.
<code>which.proteinid</code>	Use 'Protein.Accessions' (default) column for protein name. 'Master.Protein.Accessions' can be used instead to get the protein name with single protein.
<code>useNumProteinsColumn</code>	logical, TRUE (default) removes shared peptides by information of # Proteins column in PSM sheet.
<code>useUniquePeptide</code>	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
<code>rmPSM_withfewMea_withinRun</code>	TRUE (default) will remove the features that have 1 or 2 measurements within each Run.
<code>rmProtein_with1Feature</code>	TRUE will remove the proteins which have only 1 peptide and charge. Default is FALSE.
<code>summaryforMultipleRows</code>	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

data.frame of class MSstatsTMT

Examples

```
raw.pd = data.table::fread(system.file("tinytest/raw_data/PDTMT/pdtmt_input.csv",
                                     package = "MSstatsConvert"))
annotation.pd = data.table::fread(system.file("tinytest/raw_data/PDTMT/pd_annotation.csv",
                                             package = "MSstatsConvert"))
head(raw.pd)
head(annotation.pd)
```

```
input.pd <- PDtoMSstatsTMTFormat(raw.pd, annotation.pd)
head(input.pd)
```

PhilosophertoMSstatsTMTFormat

Convert Philosopher (Fragpipe) output to MSstatsTMT format.

Description

Convert Philosopher (Fragpipe) output to MSstatsTMT format.

Usage

```
PhilosophertoMSstatsTMTFormat(
  input,
  annotation,
  protein_id_col = "Protein",
  peptide_id_col = "Peptide.Sequence",
  Purity_cutoff = 0.6,
  PeptideProphet_prob_cutoff = 0.7,
  useUniquePeptide = TRUE,
  rmPSM_withfewMea_withinRun = TRUE,
  rmPeptide_OxidationM = TRUE,
  rmProtein_with1Feature = FALSE,
  summaryforMultipleRows = sum,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	data.frame of msstats.csv file produced by Philosopher
annotation	annotation with Run, Fraction, TechRepMixture, Mixture, Channel, BioReplicate, Condition columns or a path to file. Refer to the example 'annotation' for the meaning of each column. Channel column should be consistent with the channel columns (Ignore the prefix "Channel ") in msstats.csv file. Run column should be consistent with the Spectrum.File columns in msstats.csv file.
protein_id_col	Use 'Protein'(default) column for protein name. 'Master.Protein.Accessions' can be used instead to get the protein ID with single protein.
peptide_id_col	Use 'Peptide.Sequence'(default) column for peptide sequence. 'Modified.Peptide.Sequence' can be used instead to get the modified peptide sequence.
Purity_cutoff	Cutoff for purity. Default is 0.6

PeptideProphet_prob_cutoff	Cutoff for the peptide identification probability. Default is 0.7. The probability is confidence score determined by PeptideProphet and higher values indicate greater confidence.
useUniquePeptide	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
rmPSM_withfewMea_withinRun	TRUE (default) will remove the features that have 1 or 2 measurements within each Run.
rmPeptide_OxidationM	TRUE (default) will remove the peptides including oxidation (M) sequence.
rmProtein_with1Feature	TRUE will remove the proteins which have only 1 peptide and charge. Default is FALSE.
summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing will be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If append = TRUE, has to be a valid path to a file.
...	additional parameters to <code>data.table::fread</code> .

Value

data.frame of class MSstatsTMT

Examples

```
input_file_path = system.file("tinytest/raw_data/Philosopher/msstats.csv",
                             package = "MSstatsConvert")
annotation_file_path = system.file("tinytest/raw_data/Philosopher/MSstatsTMT_annotation.csv",
                                   package = "MSstatsConvert")
input = data.table::fread(input_file_path)
annotation = data.table::fread(annotation_file_path)
msstats_format = PhilosophertoMSstatsTMTFormat(input, annotation)
head(msstats_format)
```

 ProgenesistoMSstatsFormat

Import Progenesis files

Description

Import Progenesis files

Usage

```
ProgenesistoMSstatsFormat(
  input,
  annotation,
  useUniquePeptide = TRUE,
  summaryforMultipleRows = max,
  removeFewMeasurements = TRUE,
  removeOxidationMpeptides = FALSE,
  removeProtein_with1Peptide = FALSE,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	name of Progenesis output, which is wide-format. 'Accession', 'Sequence', 'Modification', 'Charge' and one column for each run are required.
annotation	name of 'annotation.txt' or 'annotation.csv' data which includes Condition, BioReplicate, Run information. It will be matched with the column name of input for MS runs.
useUniquePeptide	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
removeFewMeasurements	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeOxidationMpeptides	TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.

```

removeProtein_with1Peptide
    TRUE will remove the proteins which have only 1 peptide and charge. FALSE
    is default.
use_log_file    logical. If TRUE, information about data processing will be saved to a file.
append         logical. If TRUE, information about data processing will be added to an existing
log file.
verbose        logical. If TRUE, information about data processing will be printed to the con-
sole.
log_file_path  character. Path to a file to which information about data processing will be
saved. If not provided, such a file will be created automatically. If append =
TRUE, has to be a valid path to a file.
...           additional parameters to data.table::fread.

```

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek, Ulrich Omasits

Examples

```

progenesis_raw = system.file("tinytest/raw_data/Progenesis/progenesis_input.csv",
                             package = "MSstatsConvert")
annot = system.file("tinytest/raw_data/Progenesis/progenesis_annot.csv",
                   package = "MSstatsConvert")
progenesis_raw = data.table::fread(progenesis_raw)
annot = data.table::fread(annot)

progenesis_imported = ProgenesisToMSstatsFormat(progenesis_raw, annot,
                                                use_log_file = FALSE)

head(progenesis_imported)

```

ProteinProspectorToMSstatsTMTFormat

Generate MSstatsTMT required input format from Protein Prospector output

Description

Generate MSstatsTMT required input format from Protein Prospector output

Usage

```

ProteinProspectortoMSstatsTMTFormat(
  input,
  annotation,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeProtein_with1Feature = FALSE,
  summaryforMultipleRows = sum,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL
)

```

Arguments

<code>input</code>	Input txt peptide report file from Protein Prospector with "Keep Replicates", "Mods in Peptide", and "Protein Mods" options selected.
<code>annotation</code>	data frame which contains column Run, Fraction, TechRepMixture, Mixture, Channel, BioReplicate, Condition.
<code>useUniquePeptide</code>	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
<code>removeFewMeasurements</code>	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
<code>removeProtein_with1Feature</code>	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
<code>summaryforMultipleRows</code>	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.

Value

data.frame of class "MSstatsTMT"

Examples

```
input = system.file("tinytest/raw_data/ProteinProspector/Prospector_TotalTMT.txt",
  package = "MSstatsConvert")
input = data.table::fread(input)
annot = system.file("tinytest/raw_data/ProteinProspector/Annotation.csv",
  package = "MSstatsConvert")
annot = data.table::fread(annot)
output <- ProteinProspectortoMSstatsTMTFormat(input, annot)
head(output)
```

SkylinetoMSstatsFormat

Import Skyline files

Description

Import Skyline files

Usage

```
SkylinetoMSstatsFormat(
  input,
  annotation = NULL,
  removeiRT = TRUE,
  filter_with_Qvalue = TRUE,
  qvalue_cutoff = 0.01,
  useUniquePeptide = TRUE,
  removeFewMeasurements = TRUE,
  removeOxidationMpeptides = FALSE,
  removeProtein_with1Feature = FALSE,
  use_log_file = TRUE,
  append = FALSE,
  verbose = TRUE,
  log_file_path = NULL,
  ...
)
```

Arguments

input	name of MSstats input report from Skyline, which includes feature-level data.
annotation	name of 'annotation.txt' data which includes Condition, BioReplicate, Run. If annotation is already complete in Skyline, use annotation=NULL (default). It will use the annotation information from input.
removeiRT	TRUE (default) will remove the proteins or peptides which are labeled 'iRT' in 'StandardType' column. FALSE will keep them.

<code>filter_with_Qvalue</code>	TRUE(default) will filter out the intensities that have greater than <code>qvalue_cutoff</code> in <code>DetectionQValue</code> column. Those intensities will be replaced with zero and will be considered as censored missing values for imputation purpose.
<code>qvalue_cutoff</code>	Cutoff for <code>DetectionQValue</code> . default is 0.01.
<code>useUniquePeptide</code>	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
<code>removeFewMeasurements</code>	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
<code>removeOxidationMpeptides</code>	TRUE will remove the peptides including 'oxidation (M)' in modification. FALSE is default.
<code>removeProtein_with1Feature</code>	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

`data.frame` in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek

Examples

```
skyline_raw = system.file("tinytest/raw_data/Skyline/skyline_input.csv",
                          package = "MSstatsConvert")
skyline_raw = data.table::fread(skyline_raw)
skyline_imported = SkylinetoMSstatsFormat(skyline_raw)
head(skyline_imported)
```

SpectroMineToMSstatsTMTFormat
Import data from SpectroMine

Description

Import data from SpectroMine

Usage

```
SpectroMineToMSstatsTMTFormat(  
  input,  
  annotation,  
  filter_with_Qvalue = TRUE,  
  qvalue_cutoff = 0.01,  
  useUniquePeptide = TRUE,  
  rmPSM_withfewMea_withinRun = TRUE,  
  rmProtein_with1Feature = FALSE,  
  summaryforMultipleRows = sum,  
  use_log_file = TRUE,  
  append = FALSE,  
  verbose = TRUE,  
  log_file_path = NULL,  
  ...  
)
```

Arguments

input	data name of SpectroMine PSM output. Read PSM sheet.
annotation	data frame which contains column Run, Fraction, TechRepMixture, Mixture, Channel, BioReplicate, Condition. Refer to the example 'annotation.mine' for the meaning of each column.
filter_with_Qvalue	TRUE(default) will filter out the intensities that have greater than qvalue_cutoff in EG.Qvalue column. Those intensities will be replaced with NA and will be considered as censored missing values for imputation purpose.
qvalue_cutoff	Cutoff for EG.Qvalue. default is 0.01.
useUniquePeptide	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
rmPSM_withfewMea_withinRun	TRUE (default) will remove the features that have 1 or 2 measurements within each Run.
rmProtein_with1Feature	TRUE will remove the proteins which have only 1 peptide and charge. Default is FALSE.

summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
use_log_file	logical. If TRUE, information about data processing will be saved to a file.
append	logical. If TRUE, information about data processing will be added to an existing log file.
verbose	logical. If TRUE, information about data processing will be printed to the console.
log_file_path	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If append = TRUE, has to be a valid path to a file.
...	additional parameters to <code>data.table::fread</code> .

Value

data.frame of class MSstatsTMT

Examples

```
raw.mine = data.table::fread(system.file("tinytest/raw_data/SpectroMine/spectromine_input.csv",
                                         package = "MSstatsConvert"))
annotation.mine = data.table::fread(system.file("tinytest/raw_data/SpectroMine/spectromine_annotation.csv",
                                                package = "MSstatsConvert"))

head(raw.mine)
head(annotation.mine)
input.mine <- SpectroMinetoMSstatsTMTFormat(raw.mine, annotation.mine)
head(input.mine)
```

SpectronauttoMSstatsFormat

Import Spectronaut files

Description

Import Spectronaut files

Usage

```
SpectronauttoMSstatsFormat(
  input,
  annotation = NULL,
  intensity = "PeakArea",
  peptideSequenceColumn = "EG.ModifiedSequence",
  heavyLabels = NULL,
  excludedFromQuantificationFilter = TRUE,
```

```

filter_with_Qvalue = FALSE,
qvalue_cutoff = 0.01,
useUniquePeptide = TRUE,
removeFewMeasurements = TRUE,
removeProtein_with1Feature = FALSE,
summaryforMultipleRows = max,
calculateAnomalyScores = FALSE,
anomalyModelFeatures = c(),
anomalyModelFeatureTemporal = c(),
removeMissingFeatures = 0.5,
anomalyModelFeatureCount = 100,
runOrder = NULL,
n_trees = 100,
max_depth = "auto",
numberOfCores = 1,
use_log_file = TRUE,
append = FALSE,
verbose = TRUE,
log_file_path = NULL,
...
)

```

Arguments

input	name of Spectronaut output, which is long-format. ProteinName, PeptideSequence, PrecursorCharge, FragmentIon, ProductCharge, IsotopeLabelType, Condition, BioReplicate, Run, Intensity, F.ExcludedFromQuantification are required. Rows with F.ExcludedFromQuantification=True will be removed.
annotation	name of 'annotation.txt' data which includes Condition, BioReplicate, Run. If annotation is already complete in Spectronaut, use annotation=NULL (default). It will use the annotation information from input.
intensity	Intensity column to use. Accepts legacy enum values 'PeakArea' (default, uses F.PeakArea), 'NormalizedPeakArea' (uses F.NormalizedPeakArea). Can also be any raw Spectronaut column name passed as a string (e.g. "FG.MS1Quantity"); the column name is standardized internally. For protein turnover workflows the recommended default is "FG.MS1Quantity".
peptideSequenceColumn	Name of the Spectronaut column that contains the peptide sequence. Defaults to "EG.ModifiedSequence". The value is standardized internally (dots and spaces removed) before column lookup.
heavyLabels	Character list identifying the heavy isotope labels as it appears inside square brackets in the peptide sequence column, e.g. c("Lys6") matches peptides containing [Lys6]. c("Lys6", "Arg10") matches peptides containing either [Lys6] or [Arg10]. Supports any novel label name reported by Spectronaut (e.g. "Leu6", "Phe10"). When provided, peptides are classified as heavy (IsotopeLabelType = "H"), light (IsotopeLabelType = "L"), or unlabeled (IsotopeLabelType = NA) based on its labeled sequence. When NULL (default) all peptides receive IsotopeLabelType = "L". Useful for protein turnover experiments.

excludedFromQuantificationFilter	Remove rows with F.ExcludedFromQuantification=TRUE Default is TRUE.
filter_with_Qvalue	FALSE(default) will not perform any filtering. TRUE will filter out the intensities that have greater than qvalue_cutoff in EG.Qvalue column. Those intensities will be replaced with zero and will be considered as censored missing values for imputation purpose.
qvalue_cutoff	Cutoff for EG.Qvalue. default is 0.01.
useUniquePeptide	TRUE (default) removes peptides that are assigned for more than one proteins. We assume to use unique peptide for each protein.
removeFewMeasurements	TRUE (default) will remove the features that have 1 or 2 measurements across runs.
removeProtein_with1Feature	TRUE will remove the proteins which have only 1 feature, which is the combination of peptide, precursor charge, fragment and charge. FALSE is default.
summaryforMultipleRows	max or sum - when there are multiple measurements for certain feature and certain run, use highest or sum of multiple intensities. Default is max for label-free converters and sum for TMT converters.
calculateAnomalyScores	Default is FALSE. If TRUE, will run anomaly detection model and calculate anomaly scores for each feature. Used downstream to weigh measurements in differential analysis.
anomalyModelFeatures	character vector of quality metric column names to be used as features in the anomaly detection model. List must not be empty if calculateAnomalyScores=TRUE.
anomalyModelFeatureTemporal	character vector of temporal direction corresponding to columns passed to anomalyModelFeatures. Values must be one of: mean_decrease, mean_increase, dispersion_increase, or NULL (to perform no temporal feature engineering). Default is empty vector. If calculateAnomalyScores=TRUE, vector must have as many values as anomalyModelFeatures (even if all NULL).
removeMissingFeatures	Remove features with missing values in more than this fraction of runs. Default is 0.5. Only used if calculateAnomalyScores=TRUE.
anomalyModelFeatureCount	Feature selection for anomaly model. Anomaly detection works on the precursor-level and can be much slower if all features used. We will by default filter to the top-100 highest intensity features. This can be adjusted as necessary. To turn feature-selection off, set this value to a high number (e.g. 10000). Only used if calculateAnomalyScores=TRUE.
runOrder	Temporal order of MS runs. Should be a two column data.table with columns Run and Order, where Run matches the run name output by Spectronaut and Order is an integer. Used to engineer the temporal features defined in anomaly-ModelFeatureTemporal.

<code>n_trees</code>	Number of trees to use in isolation forest when calculateAnomalyScores=TRUE. Default is 100.
<code>max_depth</code>	Max tree depth to use in isolation forest when calculateAnomalyScores=TRUE. Default is "auto" which calculates depth as $\log_2(N)$ where N is the number of runs. Otherwise must be an integer.
<code>numberOfCores</code>	Number of cores for parallel processing anomaly detection model. When > 1, a logfile named 'MSstats_anomaly_model_progress.log' is created to track progress. Only works for Linux & Mac OS. Default is 1.
<code>use_log_file</code>	logical. If TRUE, information about data processing will be saved to a file.
<code>append</code>	logical. If TRUE, information about data processing will be added to an existing log file.
<code>verbose</code>	logical. If TRUE, information about data processing will be printed to the console.
<code>log_file_path</code>	character. Path to a file to which information about data processing will be saved. If not provided, such a file will be created automatically. If <code>append = TRUE</code> , has to be a valid path to a file.
<code>...</code>	additional parameters to <code>data.table::fread</code> .

Value

data.frame in the MSstats required format.

Author(s)

Meena Choi, Olga Vitek

Examples

```
spectronaut_raw = system.file("tinytest/raw_data/Spectronaut/spectronaut_input.csv",
                             package = "MSstatsConvert")
spectronaut_raw = data.table::fread(spectronaut_raw)
spectronaut_imported = SpectronauttoMSstatsFormat(spectronaut_raw, use_log_file = FALSE)
head(spectronaut_imported)
```

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